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### ALLEVIATION SALINITY STRESS ON TOMATO PLANTS BY SOME ORGANIC AND BIO - FERTILIZERS APPLICATION

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**ABSTRACT:** A pot experiment was carried out in two summer seasons of 2020 and 2021, to study the effect of organic {compost (COM) and humic acid (HA)} and biological {arbuscular mycorrhizal (AM) and plant growth promotion rizobacteria (PGPB)} fertilization on mitigation salinity hazard of tomato plants. Saline solutions were prepared by using NaCl to induce an EC of 3 and 6 dSm<sup>-1</sup>, in addition to tap water (0.56 dSm<sup>-1</sup>) as a control. Data on plant growth and development, and leaf water, mineral and chemical contents, and fruit yield and quality were determined. Comparable to un-saline treatment (tap water), salinity (at 3.00 and 6.00 dSm<sup>-1</sup>) decreased plant growth, fruit set (%), water, and mineral nutrition contents in leaves, as well as fruit yield. However, salinity increased water use efficiency, leaf proline content, electrolyte leakage in leaves, and fruit contents of TSS and Vit.C. Also, salinity enhanced Na and Cl contents in all leaves, particularly old ones. Treatments of alleviation salinity all mitigated salinity detrimental effect as they enhanced growth, fruit set (%), water, N, P, K and Ca contents in leaves as well as fruit yield. Na and Cl contents in both young and old leaves particularly in former ones, beside leaf proline content and leaf electrolyte leakage. The combined treatments i.e., AM+PGPR and COM+HA both seems to be of a synergistic effect as they were the most effective treatments in terms of alleviation salinity hazards on plants followed by AM and COM applied alone.

**Kew words**: Tomato, salinity alleviation treatments, plant growth, chemical contents and fruit yield, organic and bio-fertilizers.

#### INTRODUCTION

The continuous increase in the earth's human population, including the developing countries of the Mediterranean region, requires increasing quantities of water for domestic, industrial and agricultural needs. The progressive requirement for more water to irrigate crops for food when water resources are limited has led to use low quality water for irrigation, such as saline field drainage or brackish water, etc. Irrigation with saline water has become necessary in parts of the world with limited supplies of good quality water.

According to Gama *et al.* (2007), plants grown under salinity conditions are basically stressed in three ways. These are, (1) osmotic effect; reduction of water potential in the root zone and causing water deficit, i.e. excess salts in the root zone hinder roots from withdrawing water from surrounding soil, (2) specific ion effect; phototoxicity of ions such as Na<sup>+</sup> and Cl<sup>-</sup>, and (3) nutrient imbalance by depression ion uptake. Therefore, salinity stress involves changes in various physiological and metabolic processes, depending on severity and duration of the stress, and ultimately inhibits crop growth and production (Rozema and Flowers, 2008, Rahnama et al., 2010 and James et al., 2011). Osmotic stress causes various physiological changes, impairs the ability to detoxify reactive oxygen species (ROS), decreased photosynthetic activity, and decrease in stomatal aperture (Munns and Tester 2008 and Rahnama et al., 2010). Also, salinity altered the mineral nutrient composition by decreasing N,P,K and Ca content and increased Na and Cl content of the tomato plants compared to the unsalted control (Tartoura et al., 2014 and Ors et al., 2021). The accumulation of proline in plants (Ali and Rab, 2017 and Torre-Gonzalez et al., 2018), and increasing electrolyte leakage from plasma membranes proportionally in tomato leaves has been observed (Tartoura et al., 2014., and Ors et al., 2021) under salinity stress conditions. As a result, several studies showed that tomato plant growth was reduced by salinity (Feigin et al., 1987 and Magan et al., 2008), as well tomato yield is quite sensitive to salinity, i.e., at 3.0 dS m<sup>-1</sup> and above (Malash et al., 2012, El-Mogy et al., 2018 and Pengfei et al., 2019). While there was a clear reduction in yield, the fruit quality of tomato fruit (in most cases) including TSS and vitamin C., were enhanced with increasing salinity (Mizrahi et al., 1988, De Pascale et al., 2001, Malash et al., 2002 and Maggio et al., 2004). Consequently, great effort has been devoted to overcome the deleterious effects of salinity on crop plants. Biofertilizers such as arbuscular mycorrhizal (AM) and plant growth promoting rhizobacteria (PGPR) were mentioned to be have alleviation effect of salt stress on crop plants. The symbiotic association of crop plants with AM fungi improves the uptake of almost essential nutrients by plants (Balliu et al., 2015), Whereas decrease the uptake of Na and Cl (Evelin et al., 2012), In addation AM increases water uptake bymaize plant roots (Ruiz-Lozano and Azcon, 1995 and Marulanda et al., 2003). reduced electrolyte leakage in plant leaves (Ahmad et al., 2019 and Kaya et al., 2009). PGPB treatment can directly fixing atmospheric nitrogen, producing some phytohormones, solubilizing minerals such as phosphorus and synthesizing enzymes that can modulate plant growth and development (Mayak et al., 2004a). Furthermore, PGPR reduced salt toxicity in various plants by lowering the Na<sup>+</sup> concentration and increasing the K<sup>+</sup> concentration in crop plants (Bano and Fatima, 2009 and Kohler et al., 2009). The combined treatment of both mycorrhiza and PGPR seems to be has a synergistic effect that was confirmed by improved plant growth, nutrition, and yield as well as mitigated salinity stress than using one component of them alone (Baradar et al., 2015, Calvo-Polanco et al., 2016 and Desai et al., 2020).

Application of composted organic matter (OM) leads to improve soil physical, chemical and biological properties, increasing soil water-holding capacity and bulk density and improving plant nutrient use efficiency (Qadir and Oster 2004, Tejada *et al.*, 2006, Clark *et al.*, 2007, and Altome *et al.*, 2015). Application of compost increased the N, Ca, P, K, Mg, Fe, Zn, and Cu contents in plants grown under saline conditions

(Dursun et al., 2002, and Du Jardin, 2015), while it reduces the uptake of some toxic elements (Knicker et al., 1993, and Friedel and Scheller, 2002), and reduces electrolyte leakage (EL) in plants that were grown in saline soil (Rady et al., 2016). Regarding, humic acid (HA) it was able to stimulate nutrient uptake such as N, Ca, P, K, Mg, Fe, Zn, and Cu (Padem et al., 1997, and Dursun et al., 2002), and their use efficiency by plants, meanwhile reduced the uptake of some toxic elements (Knicker et al., 1993, and Friedel and Scheller, 2002). Also, HA improved RWC in strawberry plants (Saidimoradi et al., 2019) significantly reduced electrolyte leakage in bean (Phaseolus vulgaris L.) plants (Aydin et al., 2012), besides decreasing membrane damage (Canellas et al., 2015) which can mitigate the deleterious effects of salt stress (Du Jardin, 2015).

This study was undertaken to provide information about the possibility of organic and bio-fertilizers in enhancing salt tolerance in plants, thus we hypothesized that AM, PGPR, COM and HA can alleviate salinity hazard in tomato plants grown under saline conditions.

#### MATERIALS AND METHODS

A pot experiment was carried out in two successive years in early summer seasons of 2020 and 2021, under protected conditions (theram house), at the Agricultural Experimental farm, Faculty of Agriculture Menofia University Shebin EL-Kom, Egypt. This experiment was conducted to study the effect of two sources of fertilizers; i.e., organic and biological fertilizers on reducing salinity hazard on tomato.

In this study, seeds of tomato "hybrid 186" were sown in seedling trays (209 holes) on the 10<sup>th</sup> and 8<sup>th</sup> of February in 2020 2021 yearsears, respectively. The trays were filled with a mixture of peat moss, vermiculite and mineral nutrients. The seedlings were transplanted (45 days after seed sowing) in perforated plastic pots 35cm in diameter, under theram house conditions. Each pot contained 12kg mixture of field soil and washed sand (1: 2 by weight), some washed gravels (with different sizes) were added at the bottom of each pot to optimize the leaching process. The experiment was designed in a split

plot design with 6 replicates. Each subplot consisted of 6 pots and each pot contained 5 seedlings. Salinity treatments (3 levels) were devoted to main plots whereas, fertilization sources treatments were devoted to the sub- plots. Unless otherwise indicated, fertilizers rates were added as commonly used in tomato production field i.e, 120 unit of N (600kg /fed as ammonium sulphate), 50 units of P (320 kg/fed as calcium super phosphate), and 150 units of K (300kg/fed as potassium phosphate). In addition micro – elements (iron –zinc –manganese) at a rate of 1-2g per liter of water were applied as spray on plant foliages, after a month of transplanting and repeated three times every 15 days thereafter.

At the beginning all pots were irrigated with fresh water, while salinity treatments started 20 days after transplanting. Saline solutions were prepared by using NaCl to induce EC equal to 3 and 6 dSm<sup>-1</sup>, in addition to tap water  $(0.56 \text{ dSm}^{-1})$ as a control. To avoid salinity chock, saline irrigation water was applied gradually; i.e., 2 dSm<sup>-</sup> <sup>1</sup> every 3 days till final concentration. Moisture content of pots was determined by weigh pots at 2 days intervals and irrigation was applied when soil moisture depleted to 70% of field capacity, the amount of irrigation water added was enough to raise moisture to 100% field capacity. In addition, excess of water (15% as leaching fraction) was also applied (if needed). After each irrigation the drain water was gathered in the dish below each pot and its EC was determined. The 15% leaching fraction was sufficient to keep salinity level in drain solution as in irrigation one, under condition of this experiment.

#### Salinity alleviation treatments were

#### **1-Biological fertilizers**

#### 1-1- Endo-Mycorrhizae, (Arbuscular mycorrhizal) (AM)

The fungus was added (before transplanting) to the soil in each pot at rate of 1g/kg soil, and mixed thoroughly with the soil surface.

#### 1-2-Plant growth promoting rhizobacteria (PGPR)

Roots of seedlings before transplanting were dipped in the *Bacillus subtilis\_suspension* of 10<sup>8</sup>

CFU ml<sup>-1</sup> for 5 min amended with Arabic gum solution (1%) as a sticker.

## 1-3- Mycorrhiza + Plant growth promoting rhizobacteria

Both of them (as a bilateral treatment) were added at dates previously mentioned for each, and with the same quantities.

#### 2- Organic fertilizers

#### 2-1-Compost (COM)

Compost contains 1% nitrogen, it was added before planting in a rate of 6 ton/fed<sup>-1</sup>, which is consider only as 50% of the necessary nitrogen needed for tomato production fields. The compost was mixed well in the surface layer of the potted soil.

#### 2-2-Humic acid (HA)

Humic acid "Agro Master" is a water soluble potassium humate crystals (K<sub>2</sub>O) 10% W/W, Humic acid was added at 15- 20 days after transplanting at a rate of 1 kg /fed <sup>-1</sup> (0.01 g / pot), and the application was done every 2 weeks during the growing season.

#### 2-3- Compost + humic acid

Compost and humic acid (as a bilateral treatment) were added at the dates previously mentioned for each material and with the same quantities.

#### **Data recorded**

#### I. Vegetative growth characters

A plant sample was taken at 50 days after transplanting (after three weeks of reaching the final concentration of salts) in both seasons of study, whereas in the  $2^{nd}$  season two plant samples were taken; at 50 and 60 days after transplanting (DAT). The sample consisted of 2 plants from each replicate (pot), then the following measurements were recorded:

- **1-Plant height**: was measured from cotyledon leaves scar to terminal bud.
- 2- Total plant dry weight: dry weight was determined by put all the plant organs in an oven at 70  $^{\circ}$  till constant weight.

#### **II-** Flowering date and fruit setting

- 1- Flowering date (F<sub>50</sub>): is the date (number of days) at which 50% of plants produce the first flower.
- **2- Fruit set (%):** flowers of the 3<sup>rd</sup> and 4<sup>th</sup> clusters were tagged and fruits that set were calculated.

#### **III-** Plant water relations

**1-Relative water content (RWC)**: the 5<sup>th</sup> leaf from the plant top were taken from three randomly selected plants from each treatment at 50 day after transplanting in both seasons of study. The RWC was calculated by the following equation as cited after Barrs and weatherly (1962).

$$RWC = \frac{FW - DW}{TW - DW} \times 100$$

Where: **FW**= fresh weight of leaflet.

**DW**= dry weight of leaflet (leaflets were dried up in an oven at  $70^{\circ}$ C till constant weight)

**TW**= full-turgor weight; i.e., turgor weight was determined by floated leaflet on distilled water in for 6h petri dishes under laboratory conditions, and then weighed every 15 minutes. At constant weight, leaflets were got out of the water and were blotted before reweighing.

**2-Water use efficiency (WUE)**: It was measured at the end of the season according to the following formula: **WUE**=Total Yield (kg)/ *Total Water /m*3

#### **IV-Chemical composition of tomato leaves**

 Mineral elements contents: Total nitrogen (N), potassium (K), phosphorus (P), calcium (Ca), sodium (Na) and chloride (Cl) were all determined in young active leaves (the 4th and 5th leaves from the tip of plants) and old leaves (7<sup>th</sup>, 8<sup>th</sup> and 9<sup>th</sup> from the tip of plants). These elements were determined at 50 days after transplanting (DAT) in 2020 season and at 50 days and 60 DAT in 2021. The methods used in their determinations were according to those mentioned by Pergel (1945) for N, Page *et al.*, (1982) for K, Ca and Cl, Chapman and Pratt (1961) for P, and Johanson and Ulrichs (1959) for Na. **2) Proline content:** Proline content was measured at 50 days after transplanting in both seasons, according to the method described by Bates *et al.* (1973).

**V- Electrolyte leakage (EL):** Was determined at 50 days after transplanting in both seasons of study. Electrolyte leakage is an index of physiological stresses which reflecting the damage of cell membranes and stability results in leakage of cell contents. Electrolyte leakage was determined as described by Sun *et al.*, (2006).

#### VI- Yield and its components

1) Average fruit weight: Was obtained by dividing total weight of the marketable fruits (from each treatment) by their number.

**2) Total yield:** was the weight of the all harvested fruits (ripe fruits were harvested every 2-3 days/week) throughout the entire harvesting season

#### VII-Fruit quality was determined in firm mature red fruits once at the harvesting

- 1) Total soluble solids content (TSS) was measured using an abbe hand Refractometer.
- 2) Ascorbic acid content in tomato juice (vitamin C): its determination was carried out using 2, 6, dichlorophenol indophenol dye and oxalic acid as extractor as described in AOAC (1995).

#### **Data Statistical analysis**

The data of the two seasons were statistically analyzed using the CoStat Package program, version 6.311(Cohort software, USA). The differences among the means of treatments were tested using the least significant differences (L.S.D) at 0.05 level of probability according to the method described by Snedecor and Cochran (1980).

### RESULTS AND DISCUSSIONS

### 1-Plant vegetative growth

#### 1-1- Plant height

Data in Table 1 show that increasing salinity level significantly decreased plant height of

tomato plants compared to those of un-salinized plants, in both sampling dates and seasons. The proportion of the reduction in plant height also was more pronounce by increasing time of exposure to salinity; i.e., at 60 DAT than at 50 DAT in 2021 season. Similar results were obtained by Malash *et al.* (2008), Oztekin and Tuzel (2011) and El-Mogy *et al.* (2018) who reported that salinity stress reduces the height of tomato plants. The reduction in plant height by salinity was mainly due to reduce water potential, which causes ion imbalance and ion toxicity (Gama *et al.*, 2007, Rahnama *et al.*, 2010 and James *et al.*, 2011).

Concerning, the effect of salinity alleviation treatments. Table (1) shows that all these treatments increased significantly plant height than that of the untreated control, in both years of study. Also, the most effective treatment in alleviation salinity's detrimental effect on the stem length of tomato plants was combined COM and HA in both seasons. The second highest value was recorded to plants treated with COM, followed by those treated by the combination of AM+PGPR treatment in 2020 season, me,anwhile the differences between these two particular treatments were not significant (Table 1).

Table (1): Effect of salinity levels (A), some salinity alleviation treatments (B) and their interactions(A×B) on plant height of tomato plants determined at 50d (in 2020 & 2021) and 60d (in2021 only) after transplanting.

			Salinity a	alleviation tr	reatments (I	<b>B</b> )		
Salinity loyals			]	plant height	(cm)			
dS/m		Sa	mple taken at	50 d after t	ransplantin	g in 2020		
(A)	Mycorrhizal Inoculation	B. subtilis Inoculation	Mycorrhizal + B. subtilis	Compost Application	Humic Acid Application	Compost + Humic Acid	Untreated Control	Mean A
0.56*	71.40	68.71	74.46	76.33	70.58	76.58	66.63	72.10
3.00	68.67	67.25	69.17	69.25	67.92	74.17	60.58	68.14
6.00	66.17	62.50	66.25	66.71	65.83	69.75	55.00	64.60
Mean B	68.74	66.15	69.96	70.71	68.11	73.50	60.74	
L.S.D A				0.390				
L.S.D B				0.596				
L.S.D AxB				1.032				
		1 <sup>st</sup> samp	Seaso Seaso	n 2021 ) d after trai	nsplanting			
0.56*	69.00	58.25	73.25	68.25	67.25	82.75	50.75	67.07
3.00	60.66	56.25	64.75	63.50	59.75	67.25	44.00	59.45
6.00	53.50	52.50	56.25	52.50	53.00	59.50	37.50	52.11
Mean B	61.06	55.67	64.75	61.42	60.00	69.83	44.08	
L.S.D A				1.898				
L.S.D B				2.899				
L.S.D A x B				5.022				
		2 <sup>nd</sup> samp	Seaso ble taken at 60	n 2021 ) d after trai	nsplanting			
0.56*	70.18	65.68	74.75	71.00	67.63	84.50	52.25	69.43
3.00	62.25	57.25	65.88	64.38	60.00	68.55	43.65	60.28
6.00	54.00	53.00	57.50	54.25	53.33	60.13	39.25	53.06
Mean B	62.14	58.64	66.04	63.21	60.32	71.06	45.05	
L.S.D A				1.110				
L.S.D B				1.695				
L.S.D A x B				2.936				

\*= tap water (control)

However, using B.subtilis (which belongs to PGPR group) alone gave the lowest value of plant height among the other treatments (Table 1). These results are similar to the data obtained in an earlier studies regarding to the favorable effect of these salinity alleviation treatments on stem length of tomato plants, e.g., Basak et al. (2011) and Hadad et al. (2012) for using AM, Tank and Saraf (2010), Pandy and Gapta (2020) and Yilmaz et al. (2020) for using PGPR, Arancon et al. (2003) and Tu et al. (2006) for using compost and Ashraf and Mohamed (2008) and Feleafel and Mirdad (2014a), for using HA. The favorable effect of AM on plant height of tomato plants was mainly attributed to its efficiency in providing nutrition to the host plant, increasing water uptake, production of hormones and enhancing adaptation to environmental stress including salinity (Garg and chandel 2010). Moreover, PGPR as well, enhances biological N2 fixation. increasing the availability of nutrients in the rhizosphere (Glick, 2012 and Vessey, 2003). In addition, Richardson et al. (2009) and Glick, (2012) added that PGPR can enhancing other beneficial symbioses of the host such as inhibition of cell wall-degrading enzymes, lowering plants ethylene levels, by which abiotic stress tolerance increased in plants.

It was also found that compost can improve soil fertility and increase the crop accessibility to nutrients, leading to good plant growth as well as reducing the damaging effects of salt stress (Cimrin *et al.*, 2010) on pepper. HA application also, has favorable effect on plants under stress conditions, as it increased nutrients uptake, (Adani *et al.*, 1998 and Dursun *et al.*, 2002), changing ion balance, promoting plasma membrane proton pumps activity and enhancing photosynthesis of tomato plants grown under salt stress (Souza *et al.*, 2021).

The highest value of plant height of tomato plants (Table1) was obtained by the combined treatment of COM+HA at 0.56 dS/m level of salinity i.e. 76.6, 82.7 and 84.5 cm at 50 DAT in 2020 and 2021 seasons and at 60 DAT in 2021 respectively, however the lowest values were

55.0, 37.5 and 39.25 in the same order were recorded to the untreated control with 6.0 dS/m.

#### 1-2-Total plant dry weight

It is obvious from results presented in Table 2 that salinity, (regardless salinity alleviation treatments) significantly decreased total plant dry weight with increasing salinity level in irrigation water, in both seasons and sampling dates. The reduction in total plant dry weight of tomato by salinity were 26.7 and 46.9% (as average of values obtained in the two seasons at 50 DAT) at salinity levels 3 and 6 dSm<sup>-1</sup> respectively compared to those plants irrigated with tap water. The reduction was augmented when dry weight of tomato plant was determined at 60 DAT in 2021 season as such reductions were 31.0 and 55.0 % at 3 and 6 dSm<sup>-1</sup> levels of salinity respectively (Table 2). These findings support the observations made by Cruz et al. (1990), Saranga et al., (1993), Malash et al., (2008), Eraslan et al. (2015) and El.Mogy et al. (2018) who mentioned that dry weight of tomato plants was reduced in proportion to the increase in salinity of the irrigation water. Also, De Pascale et al. (2003) found that irrigation pepper plants by saline water (EC of 4.4 dSm<sup>-1</sup>) resulted in 46% reduction in plant dry weight. The reduction in plant dry weight due to increasing salinity levels may be a result of a combination of osmotic and specific ion effects of Cl and Na on plants (Cruz et al., 1990 and Saranga et al., 1993).

Salinity alleviation treatments used in this study all resulted in a significant increase in salt tolerance of tomato plants as they enhanced total plant dry weight than those of untreated control (Table 2). Also, salinity alleviation treatments enhanced total plant dry weight in both saline and non-saline conditions. The combined treatment between COM+HA was the most effective treatment in increasing the dry weight of tomato plants, grown under saline conditions, among other treatments in both seasons and sampling dates i.e. at 50 and 60 DAT (Table 2). Also, the combined treatment of AM+PGPR gave the 2nd highest total plant dry weight in the 2021 season in both sampling dates, but the such treatment gave the 3<sup>rd</sup>. highest value of plant dry weight at 50 DAT of the 2020 season.

<b>Table (2):</b>	Effect of salinity levels (A), some salinity alleviation treatments (B) and their interactions
	(A×B) on total plant dry weight of tomato plants determined at 50 d (in 2020 & 2021) and
	60 d (in 2021only) after transplanting.

			Salinity all	eviation trea	tments (B)			
Salinity			Total pla	nt dry weigł	nt g/plant			
levels		Sa	mple taken at 50	0 d after tra	nsplanting i	n 2020		
dS/m (A)	Mycorrhizal Inoculation	B. subtilis Inoculation	Mycorrhizal + B. subtilis	Compost Application	Humic Acid Application	Compost + Humic Acid	Untreated control	Mean A
0.56*	21.19	17.42	22.03	25.18	19.90	26.28	11.14	20.45
3.00	15.87	14.20	16.95	17.97	15.25	20.32	8.69	15.61
6.00	13.31	12.48	14.30	14.89	11.56	15.91	6.05	12.64
Mean B	16.79	15.22	17.13	18.79	16.38	19.72	11.40	
L.S.D A				0.362				
L.S.D B				0.553				
L.S.D AxB				0.957				
			Season	2021				
	1	1 <sup>st</sup> san	ple taken at 50	d after trans	splanting		[	
0.56*	24.52	20.07	32.84	31.48	23.33	38.38	16.33	26.71
3.00	19.01	15.46	24.20	21.67	17.09	25.23	10.34	19.00
6.00	12.60	9.45	15.56	14.21	10.81	17.67	6.44	12.39
Mean B	18.71	14.99	22.83	20.59	18.51	25.46	12.90	
L.S.D A				1.038				
L.S.D B				1.585				
L.S.D. AxB				2.746				
			Season	2021				
		2 <sup>nd</sup> san	nple taken at 60	d after tran	splanting		r	
0.56*	35.07	29.01	41.04	38.14	32.25	47.11	21.84	34.92
3.00	23.65	20.48	29.31	27.09	21.94	32.45	13.85	24.11
6,00	16.13	13.74	18.21	16.96	15.51	20.42	9.05	15.72
Mean B	24.95	21.07	29.52	27.40	23.23	33.33	14.91	
L.S.D A				1.168				
L.S.D B				1.785				
L.S.D A xB				3.089				

\*= tap water (control)

These findings suggest that such combined treatments had a synergistic effect as values of plant dry weight obtained by these particular treatments, were higher than that obtained by each factor (one of its components) used alone (Table 2). AM inoculation and COM application treatments gave also high values of plant dry weight. Similar results were obtained by Altome *et al.* (2015) regarding the favorable effect of COM application on shoot dry weight of tomato

plants, and by Padem *et al.* (1997), Adani *et al.* (1998) and Dursun *et al.* (2002) regarding the favorable effect of HA application on tomato plant growth, which all were grown under saline conditions. The role of both organic fertilizers i.e. COM and HA in mitigation of salinity effect is to enrich the soil with organic matter and humic substances which improve soil physical, chemical and biological properties which enhances macro and micronutrients uptake (Walker and Bernal,

2008 and Wright et al., 2008) and increase moisture conservation which stimulates crop growth and quality (Zribi et al., 2011). Similarly, the synergistic effect of the combined treatment of AM+PGPR was also observed by Desai et al. (2020) who confirmed that both AM and PGPR applied together improved tomato plant growth, grown under salinity conditions than used each of them alone. Moreover, it was reported (Altunlu, 2020) that PGPR enhanced AMF positive effect which positively improved plant growth and physiological parameters of pepper plants under all studied salinity stress levels. The favorable effect of AM on plant growth particularly under saline conditions is mainly due to providing nutrients to the host plant, increasing water uptake, production of hormones and enhancing adaptation to environmental stresses (Garg and Chandel 2010). PGPR as well reduces synthesis of harmful ethylene which increases under stress conditions (Glick, 2014), fixing atmospheric nitrogen, phytohormone production, solubilizing minerals, modulate plant growth (Mayak et al., 2004a) and enhanced scavenging activities of reactive oxygen species (ROS) (Jianmin et al., 2014). It was also observed that salinity alleviation treatments increased total tomato plant dry weight in both normal and saline conditions (Table 2), but it seems that these treatments were somewhat more effective in saline than in normal conditions.

According to the data of the interaction between salinity levels and salinity alleviation treatments, Table 2 shows that the highest total plant dry weight obtained was a result of using the combined treatment of COM+HA along with 0.56 dS/m salinity level (un-saline), the 2<sup>nd</sup> highest value of total plant dry weight was recorded to the COM treatment at 50 DAT in 2020 season, but this ranked was recorded to the combined treatment of AM+PGPR at 50 and 60 DAT in 2021 season, all along with 0.56 dS/m salinity level. While, the lowest values of total plant dry weight were due to those plants subjected to the highest salinity level (6.0 dS/m) and those untreated with any of salinity alleviation treatments (Table 2). HA application and PGPR inoculation along with the highest salinity level gave also lower total plant dry weight (Table 2).

#### 2- Flowering date and fruit set

# 2-1- Number of days from transplanting to appearance of the first flower in 50 % of the plants ( $F_{50}$ )

According to the date given in Table 3, increasing salinity levels significantly decreased number of days required to first flower appearance of 50% of plants. In other words, salinity enhanced early flowering in tomato plants when compared with those grown under normal (non-saline) conditions.

Such result seems to be logical outcome as salinity dramatically affected vegetative growth, which predisposing to accelerate flowering. Similar results were obtained by Mostafizar Rahman *et al.* (2018) who found that salinity (i.e. 2 to 8 dS/m) decreased number of days required to flowering of five tomato varieties, and the effect was more pronounced with increasing salinity levels up to 8 dS/m.

Because salinity alleviation treatments improved water content, enhanced physiological and biochemical processes and reduced toxic elements in plant tissue which in turn promoting plant vegetative growth, all treatments increased No of days required to  $F_{50}$  of tomato plants (Table 3). Also, the treatments which gave highest growth parameters under saline stress, previously mentioned, also gave the longer period to  $F_{50}$ , in both seasons.

#### 2-2- Fruit set (%)

Fruit set (%) of tomato plants was significantly decreased by salinity (Table 4), and the decrease was more pronounced at 6 dS/m than at 3dS/m<sup>-1</sup> compared to those of non-saline control. These findings support the observations made by Adams and Ho, (1992) who mentioned that fruit set % of tomato was reduced by extreme salinity. The reduction in fruit set by salinity may owing to a reduction in number of flowers (Mostafizur Rahman *et al.*, 2018), or to flower loss or drop as a result of the restriction of water supply (Saito and Ito 1967) or for a reduction in potassium (Besford and Maw, 1975) and phosphorus uptake (Menary and Stalen 1976).

Table (3): Effect of salinity levels (A), some salinity alleviation treatments (B) and their interactions (A×B) on number of days from transplanting to appearance of first flower in 50% of plants (F<sub>50</sub>) in both seasons of study.

			Salinity a	alleviation ti	eatments (B	6)		
Salinity				F50 (days	s)			
levels				Season 20	20			
dS/m (A)	Mycorrizal Inoculation	B. subtilis Inoculation	Mycorrizal + B. subtilis	Compost Application	Humic Acid Application	Compost + Humic Acid	Untreated control	Mean A
0.56*	31.17	31.00	30.00	31.50	29.83	34.17	29.00	30.95
3.00	27.83	27.33	28.67	28.50	28.33	31.17	26.50	28.33
6.00	26.67	26.83	28.50	27.67	27.33	27.00	25.50	27.07
Mean B	28.56	28.39	29.06	29.22	28.50	30.78	27.00	
L.S.D A				0.284				
L.S.D B				0.434				
L.S.D AxB				0.751				
			Seas	on 2021				
0.56*	34.75	33.75	37.25	36.50	35.00	40.50	31.50	35.61
3.00	33.75	31.75	36.50	35.25	32.25	37.50	25.50	33.21
6.00	28.50	27.00	31.00	29.50	27.50	32.75	21.50	28.25
Mean B	32.33	30.83	34.92	33.75	31.58	36.92	26.17	
L.S.D A				0.985				
L.S.D B				1.505				
L.S.D A x B				2.607				

\*= tap water (control)

Table (4): Effect of salinity levels (A), some salinity alleviation treatments (B) and their interaction
(A×B) on tomato fruit set (%) of the 3 <sup>rd</sup> , and 4 <sup>th</sup> clusters in both seasons of study.

			Salinity a	lleviation tr	eatments (B	)		
Salinity				Fruit set (%	%)			
levels				Season 202	20			
dS/m (A)	Mycorrizal Inoculation	B. subtilis Inoculation	Mycorrizal + B. subtilis	Compost Application	Humic Acid Application	Compost + Humic Acid	Untreated control	Mean A
$0.56^{*}$	65.51	61.72	69.70	78.39	64.35	91.12	57.75	69.79
3.00	60.14	56.95	60.97	61.74	58.48	63.65	47.00	58.42
6.00	50.02	42.82	50.90	52.82	47.13	53.87	36.47	47.72
Mean B	58.56	53.83	60.52	64.32	56.66	69.55	47.07	
L.S.D A				0.845				
L.S.D B				1.290				
L.S.D AxB				2.235				
			Seaso	on 2021				
0.56 *	77.48	71.67	87.26	83.75	83.75	92.26	46.67	77.55
3.00	56.25	52.50	62.92	56.65	50.00	74.98	31.72	55.00
6.00	43.33	31.09	50.00	46.67	37.30	56.65	27.97	41.86
Mean B	59.02	51.75	66.73	62.36	57.02	74.63	35.45	
L.S.D A				4.817				
L.S.D B				7.359				
L.S.D A x B				12.745				

\*= tap water (control)

Salinity alleviation treatments enhanced tomato fruit set percent in plants either grown in normal or in saline conditions (Table 4). The combined treatment of COM+HA gave significantly the highest fruit set (%) of tomato plants either grown in normal cultural media i.e., non-saline (control treatment) or in both levels of salinity, in both seasons. The 2<sup>nd</sup> highest fruit set was due to COM application treatment in 2020 season and the combined treatment of AM+PGPR in 2021 one.

The favorable effect of salinity alleviation treatments on tomatoes was depend on enhancing nutrition water and mineral uptake, photosynthetic activity which improved physiological and biochemical process, such as photosynthetic activity and subsequently improve male and female gametophyte viability and increase number of clusters /plant and number of flowers in cluster, Such modifications enhanced fruit set (%) of tomato plants grown under saline conditions. These results seem to be similar to those obtained by Feleafel and Mirdad (2014a) and Ashraf and Mohamed (2008) who mentioned

that humic substances improve a number of clusters/plant and the number of flowers/clusters of tomato plants grown under saline conditions, which is reflected on fruit set improvement.

#### **3-** Plant water relations

#### 3-1- Relative water content (RWC)

As expected salinity reduced relative water content (RWC) and the reduction was more pronounced with the highest salinity level i.e. 6dS/m in both seasons (Table 5). RWC is also called relative turgidity and is perhaps the most widely accepted method of expressing the quantity of water in plant tissue (Boyer, 1969). The findings of this study are in agreement with those reported by Yurtseven et al. (2005), Eraslan et al. (2015) and Pengfei et al. (2019) who reported that RWC in tomato plants was decreased by NaCl salinity. Psarras et al. (2008) clarify that salinity in soil or in irrigation water particularly high levels reduce water uptake by plant roots and consequently reduces water potential in tomato plant tissues.

			Salinity a	lleviation tr	eatments (B	)		
Salinity			ŀ	RWC values	(%)			
levels				Season 202	20			
dS/m (A)	Mycorrizal Inoculation	B. subtilis Inoculation	Mycorrizal + B. subtilis	Compost Application	Humic Acid Application	Compost + Humic Acid	Untreated control	Mean A
0.56*	79.75	75.30	80.88	81.75	77.69	87.69	66.95	78.57
3.00	64.32	63.43	66.22	67.01	64.15	68.20	53.54	63.84
6.00	46.34	41.23	52.38	54.81	45.44	55.61	32.71	46.93
Mean B	63.47	59.99	66.49	67.86	62.43	70.50	51.06	
L.S.D A				0.843				
L.S.D B				1.289				
L.S.D AxB				2.231				
			Seaso	n 2021				
0.56*	71.95	67.45	79.83	73.58	70.58	83.66	55.90	71.85
3.00	60.65	56.13	66.08	63.33	58.21	68.12	44.14	59.52
6 .00 <sup>1</sup>	46.76	39.49	54.69	49.82	42.61	57.94	28.96	45.75
Mean B	59.79	54.36	66.87	62.24	57.14	69.91	43.00	
L.S.D A				1.173				
L.S.D B				1.792				
L.S.D A x B				3.104				

Table (5): Effect of salinity levels (A), some salinity alleviation treatments (B) and their interactions (A×B) on relative water content (RWC) in tomato leaf determined at 50 d after transplanting in both seasons of study.

\*= tap water (control)

Using salinity alleviation treatments can manage salinity hazard on the water content of tissues in tomato leaves, as they increased significantly RWC than untreated plants, also such treatments enhanced water content (RWC) in plants grown under normal conditions (nonsaline) as shown in Table 5. The most effective treatments in increasing RWC were the combined treatment of COM+HA, followed by the combined treatment of (AM+PGPR) and then the compost application treatment.

This study confirmed the previous reports regarding the enhancement of AM for water uptake by improving root water flow to colonized roots directly to plants (Koide, 1993 Marulanda et al., 2003). Also, it was observed that PGPR inoculation treatment resulted in a significant improvement of RWC in leaves of sweet pepper plants (AL-Kahtani et al. 2020) and strawberry plants (Karlidag et al., 2013) grown under saline conditions, comparable to the control (untreated). This favorable effect of bacteria treatment has been related to timprovedove root development and net water uptake in plants that suffer from salinity (Marulanda et al., 2006). The combined treatment of AM+PGPR i.e. Glomus spp + Bacillus subtilis, , resulted in enhanced RWC in both lettuce and tomato irrigated with 25 and 50 mM NaCl, rather than the control untreated, (Miceli et al. 2021). Also, COM when replace about 50% of NPK dose revealed a significant increase in RWC in bean plants grown in saline soil (Rady et al., 2016).

The improvement of HA on RWC in plants even grown under saline conditions was also reported by Saidimoradi *et al.* (2019), on strawberries and by Feleafel and and Mirdad (2014b) on tomato, compared to those of untreated plants .The role of both COM and HA in enhanced RWC may be due to the effect on increasing soil with organic matter and humic substances which improve soil physical properties in a way that improves water holding capacity and bulk density under salt stress conditions (Altome *et al.*, 2015). The highest value of RWC (Table 5) was obtained by the combined treatment of COM+HA at 0.56 dS/m level of salinity in 2020 and 2021 seasons, however the lowest values were recorded to the untreated control with 6.0 dS/m (Table 5).

#### **3-2-** Water use efficiency (WUE)

Data in Table 6 show that salinity enhanced WUE (which is: total fruit yield/water amount used throughout the season) and this effect was pronounced at 6 dS/m than 3 dS/m levels of salinity in both seasons of study. These finding are in agreement with those reported by Malash *et al.* (2008) who indicated that water use efficiency (WUE) of tomato plants was increased by using irrigation water with low and moderate salinity levels (2 and 3dS/m) as compared to those obtained with non-saline water (0.55dS/m).

Salinity alleviation treatments significantly increased WUE than those obtained by the untreated control (Table 6). Again, the combined treatments i.e. COM+HA and AM+PGPR as well as AM and COM each applied alone gave higher values of WUE (Table 6). These results seem to be in accordance with those obtained by Hajiboland et al. (2010) who found that inoculated arbuscular mycorrhizal (AM) improved WUE of tomato plants that grown under saline conditions. PGPR as well increased the WUE of tomato plants grown under saline conditions (Mayak et al., 2004b), also PGPR inoculation resulted in longer roots which might be helpful in the uptake of relatively more water even under salinity stress (Dodd et al., 2004 and Abd El-Samad et al., 2004) such conditions lead to better use efficiency. Organic fertilizer (COM+HA) also enhanced WUE by increasing water holding capacity in soil suffering from salinity (Altome et al., 2015) or maintaining better leaf water content under osmotic stress (Canellas et al., 2015). Accordingly, Feleafel and Mirdad (2014b) found that increasing HA rate led to a significant increase in WUE of tomato plants grown under salt stress conditions, than those of untreated control.

			Salinity	alleviation tr	eatments (B)			
Salinity				W.U.E. (kg/	<sup>/</sup> m <sup>3</sup> )			
levels				Season 202	20			
dS/m (A)	Mycorrizal Inoculation	B. subtilis Inoculation	Mycorrizal + B. subtilis	Compost Application	Humic Acid Application	Compost + Humic Acid	Untreated control	Mean A
$0.56^{*}$	1.97	1.77	2.05	2.05	1.84	2.17	1.31	1.88
3.00	2.20	1.95	2.37	2.46	1.96	3.05	1.60	2.23
6.00	2.62	2.46	2.93	3.18	2.69	3.30	1.53	2.67
Mean B	2.26	2.06	2.45	2.56	2.16	2.84	1.48	
L.S.D A				0.015				
L.S.D B				0.023				
L.S.D AxB				0.039				
			Sea	son 2021				
0.56*	1.27	0.90	1.43	1.34	1.12	1.72	0.82	1.23
3.00	1.50	1.25	1.78	1.60	1.43	1.97	0.92	1.49
6.00	1.76	1.35	2.49	2.07	1.58	2.66	0.93	1.84
Mean B	1.51	1.17	1.90	1.67	1.38	2.12	0.89	
L.S.D A				0.023				
L.S.D B				0.036				
L.S.D AxB				0.062				

Table (6): Effect of salinity levels (A), some salinity alleviation treatments (B) and their interactions (A×B) on water use efficiency (WUE)\*\* of tomato plants determined at the end of both seasons of study.

\*= tap water (control)

\*\*WUE= Total yield (kg) / water used throughout the growing season (m<sup>3</sup>).

#### 4-Effect on leaf chemical content

# 4-1- Mineral elements contents in young and old leaves

Salinity of irrigation water in this study resulted in decreasing tomato leaf contents of important essential nutrient elements i.e., N, P, K and Ca in both young and old leaves in both seasons, and sampling dates (Tables, 7, 8, 9 and 10). Same tables also show that the reduction of these elements was more pronounced in plant sample taken at 60 DAT than that taken at 50 DAT. It also observed that young leaves had higher levels of nutrient elements i.e., N, P and K than in old leaves (Tables, 7, 8 and 9) however Ca contents show a counter- trend i.e., old leaves had higher content of Ca than those in young leaves in both seasons and sampling dates (Table 10). On the other hand, both Na and Cl contents were found in both young and old leaves of tomato, but their contents were much higher in old leaves than in young ones in both seasons and sampling dates (Tables 11 and 12). These results agreed with former reports regarding the detrimental effect of salinity on nutrient elements uptake and contents in plant leaves such as N, P, K and Ca, while salinity resulted in increasing Na and Cl content in tomato plant tissues (Malash et al., 2008, Tartoura et al., 2014 and Ors et al., 2021). The depression of the essential nutrient mineral's contents in plant tissues by salinity may be due to the competition and antagonism between high concentration of Na and Cl ions and such minerals (Grattan and Grieve, 1999 and Tester and Davenport, 2003).

7): Effect of salinity levels (A), some salinity alleviation treatments (B) and their interactions (A×B) on nitrogen (N) content in young and	leaves $^{**}$ of tomato plants determined at 50d (in 2020 & 2021) and 60d (in 2021 only) after transplanting.
[able (7): 1	-

							Salinit	v allevia	tion treat	ments		D				
Salinity						Sample	e taken at	50 d aft	er transl	olanting	in 2020					
dS/m			N conte	int in you	ung leave	(%) Sc					Z	content i	n old lea	Ves (%)		
(Y)	MA	PGPR	MA+ PGPR	COM Applic.	HA Applic.	COM+ HA	Untreat.	Mean A	MA Inocu	PGPR	MA+ PGPR	COM Applic.	HA Applic.	COM+ HA	Untreat. control	Mean A
0.56*	2.12	1.83	2.25	2.14	1.91	2.56	1.46	2.04	1.52	1.23	1.86	1.78	1.49	1.96	1.18	1.57
3.00	1.96	1.72	2.09	2.04	1.88	2.25	1.33	1.90	1.41	1.05	1.49	1.49	1.31	1.72	1.10	1.37
6.00	1.70	1.57	1.73	1.70	1.62	1.91	1.25	1.64	1.25	1.20	1.31	1.25	1.23	1.49	0.97	1.24
Mean B	1.93	1.71	2.02	1.96	1.80	2.24	1.35		1.39	1.16	1.55	1.51	1.34	1.72	1.08	
L.S.D. A				0.0	94							0.0	84			
L.S.D. B				0.1	144							0.1	28			
L.S.D. AxB				0.5	250							0.2	22			
					1st samp	ole taken	i at 50 d a	fter tran	Isplantin	g in 2021						
0.56*	2.40	2.38	2.53	2.43	2.40	3.53	1.70	2.48	1.72	1.49	1.86	1.72	1.65	2.98	1.46	1.84
3.00	2.17	1.93	2.46	2.27	2.04	2.64	1.59	2.16	1.54	1.33	1.67	1.59	1.33	1.72	1.05	1.46
6.00	2.09	1.88	2.27	2.17	1.88	2.27	1.46	2.00	1.49	1.18	1.65	1.41	1.31	1.62	0.91	1.37
Mean B	2.22	2.06	2.42	2.29	2.11	2.81	1.59		1.59	1.33	1.72	1.58	1.43	2.11	1.14	
L.S.D. A				0.0	080							0.0	78			
L.S.D. B				0.1	121							0.1	20			
L.S.D. AxB				0.2	210							0.2	08			
					2nd sam	ple take	n at 60 d	after tra	nsplanti	ng in 202	1					
0.56*	1.86	1.70	2.27	2.04	1.83	2.35	1.54	1.94	1.25	1.18	1.41	1.33	1.20	1.65	1.10	1.30
3.00	1.67	1.65	1.78	1.72	1.59	2.04	1.49	1.71	1.25	1.15	1.41	1.33	1.18	1.57	1.07	1.28
6.00	1.65	1.57	1.72	1.70	1.57	1.86	1.33	1.63	1.20	1.10	1.25	1.23	1.10	1.49	0.91	1.18
Mean B	1.72	1.64	1.93	1.82	1.66	2.08	1.45		1.24	1.14	1.36	1.30	1.16	1.57	1.03	
L.S.D. A				0.(	660							0.0	53			
L.S.D. B				0.1	151							0.0	82			
L.S.D. AxB				0.0	262							0.1	41			
	3															

\*= tap water (control)

\*\*= Young leaves: are the 4th and 5th leaves from plant tip, while old leaves: are the 7th, 8th and 9th leaves from plant tip.

Salinity levels dS/m         Salinity levels dS/m           (A)         (A)           (A)         (A)           (A)         (A)           (B)         (A)           (B)         (A)           (B)         (A)           (B)         (A)           (A)         (A)           (B)         (A)           (B)         (A)           (B)         (A)           (B)         (A)           (A)         (A) <th>MA Inocu. 0.154 0.145 0.145 0.145 0.142 0.142 0.142 0.142 0.142</th> <th>PGPR Inocu. 0.146 0.136 0.136 0.124 0.136 0.124 0.128 0.198</th> <th>P cor MA+ PGPR PGPR 0.171 0.157 0.153 0.153 0.153 0.154 0.154 0.250</th> <th>tent in y COM COM Applic. 0.137 0.134 0.134 0.137 0.138 0.176 0.176 0.176 0.176 0.178 0.178</th> <th>nung leav           HA           HA           HA           0.130           0.130           0.126           0.126           0.127           0.128           0.120           0.126           0.126           0.127           0.128           0.129           0.120           0.120           0.120           0.120           0.120           0.120           0.126           0.128           0.128           0.128           0.129           0.128           0.128           0.129           0.126           0.126           0.126           0.128</th> <th>Sample es (%) COM + HA + HA 0.136 0.136 0.136 0.136 0.121 0.121 0.121 0.123</th> <th>Salinity taken at Untreat. control 0.120 0.120 0.120 0.060 0.099 0.138 0.138 0.138 0.138 0.138 0.138</th> <th>30 d affi 50 d affi A A 0.136 0.198 0.198 0.179 0.179 0.179 0.179</th> <th>ion treans er transpans MA Inocu. 0.119 0.116 0.104 0.104 0.167 0.138 0.158 0.158 0.158 0.158</th> <th>PGPR PGPR Inocu. 0.116 0.116 0.105 0.105 0.124 0.124 0.124 0.124</th> <th>in 2020 Peep PGPR 0.148 0.148 0.120 0.120 0.120 0.177 0.177 0.173 0.173</th> <th>CCOM COM Applic. 0.002 0.002 0.002 0.002 0.002 0.002 0.002 0.002 0.0000 0.0000 0.0000 0.0000 0.000000</th> <th>Applic. HA Applic. 0.090 0.086 0.086 0.002 0.002 0.0127 0.0109 0.0109 0.0109 0.1004</th> <th>s(%) COM + HA + HA 0.107 0.095 0.095 0.095 0.095 0.095 0.095 0.132</th> <th>Untreat. control 0.085 0.071 0.071 0.071 0.071 0.071 0.070 0.070</th> <th>Mean A A A 0.0099 0.0091 0.0138 0.1133 0.1133 0.1133</th>	MA Inocu. 0.154 0.145 0.145 0.145 0.142 0.142 0.142 0.142 0.142	PGPR Inocu. 0.146 0.136 0.136 0.124 0.136 0.124 0.128 0.198	P cor MA+ PGPR PGPR 0.171 0.157 0.153 0.153 0.153 0.154 0.154 0.250	tent in y COM COM Applic. 0.137 0.134 0.134 0.137 0.138 0.176 0.176 0.176 0.176 0.178 0.178	nung leav           HA           HA           HA           0.130           0.130           0.126           0.126           0.127           0.128           0.120           0.126           0.126           0.127           0.128           0.129           0.120           0.120           0.120           0.120           0.120           0.120           0.126           0.128           0.128           0.128           0.129           0.128           0.128           0.129           0.126           0.126           0.126           0.128	Sample es (%) COM + HA + HA 0.136 0.136 0.136 0.136 0.121 0.121 0.121 0.123	Salinity taken at Untreat. control 0.120 0.120 0.120 0.060 0.099 0.138 0.138 0.138 0.138 0.138 0.138	30 d affi 50 d affi A A 0.136 0.198 0.198 0.179 0.179 0.179 0.179	ion treans er transpans MA Inocu. 0.119 0.116 0.104 0.104 0.167 0.138 0.158 0.158 0.158 0.158	PGPR PGPR Inocu. 0.116 0.116 0.105 0.105 0.124 0.124 0.124 0.124	in 2020 Peep PGPR 0.148 0.148 0.120 0.120 0.120 0.177 0.177 0.173 0.173	CCOM COM Applic. 0.002 0.002 0.002 0.002 0.002 0.002 0.002 0.002 0.0000 0.0000 0.0000 0.0000 0.000000	Applic. HA Applic. 0.090 0.086 0.086 0.002 0.002 0.0127 0.0109 0.0109 0.0109 0.1004	s(%) COM + HA + HA 0.107 0.095 0.095 0.095 0.095 0.095 0.095 0.132	Untreat. control 0.085 0.071 0.071 0.071 0.071 0.071 0.070 0.070	Mean A A A 0.0099 0.0091 0.0138 0.1133 0.1133 0.1133
3.00	0.186	0.168	0.224	0.134 0.134 0.126	0.129	0.138 0.133	0.097	0.154 0.137	0.125 0.125 0.113	0.120 0.120 0.095	0.133	0.074	0.075 0.065	0.119 0.078	0.058	0.106
Mean B L.S.D. A	0.193	0.166	0.225	0.146	0.141	0.153	0.097		0.123	0.114	0.155	0.0	0.081	0.107	0.058	
L.S.D. B				0.0	004							0.0	03			
L.S.D. AxB	-			0.0	800							0.0	05			

Table (8): Effect of salinity levels (A), some salinity alleviation treatments (B) and their interactions (A×B) on phosphorus (P) content in young and old

\*\*=The same foot notes as indicated in Table 7.

Lable (9): El le	tect of sain aves" of ton	nty levels nato plan	(A), son its deteri	ne samu mined at	ty allevia 50 d (in	2020 &	eatmenus 2021) an	i (B) and d 60 d (i	n 2021oi	teracuor nly) after	is (A×b) • transpl	on potas anting.	sıum (K	) content	un youn	g and old
1 1 2							Salinity	alleviati	on treat	ments						
Samuty						Sample t	aken at :	50 d afte	r transp	lanting i	n 2020					
dS/m		K	content	in young	g leaves (	(mg/kg)					K cont	ent in old	I leaves (	(mg/kg)		
(V)	MA Inocu.	PGPR	MA+ PGPR	COM	Annlic	COM+ HA	Untreat.	Mean	MA	PGPR	MA+ PGPR	COM	Annlic	COM+ HA	Untreat.	Mean A
0.56*	107.22	106.86	109.48	110.32	107.44	112.14	102.41	107.98	102.15	101.58	102.53	102.92	102.15	105.08	97.94	102.05
3.00	103.49	100.98	103.65	107.46	101.84	110.54	98.87	103.83	100.88	99.04	101.43	101.36	100.26	103.93	82.47	98.48
6.00	101.72	100.56	102.53	102.94	100.90	104.88	93.43	100.99	95.51	90.94	100.18	100.45	94.68	102.02	65.98	92.82
Mean B	104.15	102.80	105.22	106.91	103.39	109.19	98.24		99.51	97.18	101.38	101.57	99.03	103.68	82.13	
L.S.D. A				0.07	4							.0	162			
L.S.D. B				0.11	3							0.0	248			
L.S.D. AxB				0.19	6							.0	429			
					1st samp	le taken	at 50 d a	after tran	nsplantin	12 in 202	1					
0.56*	105.57	105.05	110.77	108.22	105.22	115.83	104.39	107.87	102.78	101.02	104.15	103.35	102.02	104.56	100.40	102.61
3.00	103.79	102.04	108.62	106.49	102.58	111.29	97.92	104.68	100.77	90.10	101.68	101.06	100.34	103.14	68.13	95.03
6.00	100.98	91.77	105.05	101.97	100.44	106.66	83.43	98.61	79.71	71.26	98.77	84.89	76.33	100.80	58.96	81.53
Mean B	103.44	99.62	108.15	105.56	102.75	111.26	95.25		94.42	87.46	101.53	96.43	92.90	102.83	75.83	
L.S.D. A				1.04(	0							2.(	006			
L.S.D. B				1.585	6							3.(	064			
L.S.D. AxB				2.75	3							5.	307			
					2nd sam	ple taker	n at 60 d	after tra	insplanti	ng in 20%	21					
0.56*	105.05	102.78	107.62	105.92	103.36	111.21	100.80	105.25	98.93	95.51	101.27	101.19	96.67	103.63	81.50	96.96
3.00	102.39	100.59	105.57	103.63	101.37	109.46	93.43	102.35	87.81	81.50	98.93	96.82	82.95	101.26	64.10	87.63
6.00	100.34	87.31	103.19	102.78	98.77	103.35	79.13	96.41	82.47	66.78	93.43	90.94	75.39	100.29	54.79	80.59
Mean B	102.59	96.89	105.46	104.11	101.17	108.00	91.12		89.74	81.27	97.88	96.31	85.01	101.73	66.80	
L.S.D. A				0.68	4							1.	489			
L.S.D. B				1.04	4							2	275			
L.S.D. AxB				1.80	6							3.5	940			

d ald • . C.E. . Â ~ .... . • . d the é : -Ě . . 1.4. ÷ . -. 17.00 00 Table

\*= tap water (control)

\*\*= The same foot notes as indicated in Table 7.

1 able (10): E 0	lect of	salunty I s** of tom	evels (A) tato plan	) and som its determ	e salınıty iined at 5	alleviat 0 d (in 2	100 UreaU 1020 & 20	nents (1 21) and	9 and th 60 d (in	eır inter 2021only	actions ( y) after ti	A×B) on ransplant	calcium ( ling.	(Ca) con	tent in yo	ung and
Collination							Salinity	v allevia	tion trea	tments						
Javale						Sample	e taken at	50 d afi	er trans	planting	in 2020					
dS/m		Ü	a conten	t in young	g leaves (1	ng/kg)					Ca	content in	n old leav	es (mg/k	g)	
(V)	MA	PGPR	MA+	COM	ΗA	COM	Untreat.	Mean	MA	PGPR	MA+	COM	HA	COM	Untreat.	Mean
(2)	Inocu.	Inocu.	PGPR	Applic.	Applic.	+ HA	control	A	Inocu.	Inocu.	PGPR	Applic.	Applic.	+ HA	control	A
0.56*	8.75	7.17	9.75	9.25	8.08	12.08	6.33	8.77	13.75	12.17	16.33	14.75	13.08	16.50	9.75	13.76
3.00	7.58	6.50	8.92	8.42	7.08	10.00	5.00	7.64	12.33	11.25	14.08	13.33	11.25	14.92	7.50	12.10
6.00	5.25	5.42	6.33	5.92	4.42	7.33	3.33	5.43	9.58	7.58	11.58	12.42	8.83	12.25	4.83	9.58
Mean B	7.19	6.36	8.33	7.86	6.53	9.81	4.89		11.89	10.33	14.00	13.50	11.06	14.56	7.36	
L.S.D. A				0.5	572							0.7	731			
L.S.D. B				0.8	14							1.1	17			
L.S.D. AxB				1.5	14							1.5	35			
					1st sam	ple take	en at 50 d	after tr	unsplant	ing in 20	21					
0.56*	9.25	7.50	10.50	9.67	8.83	12.50	6.50	9.25	13.67	12.08	16.33	15.33	12.67	17.25	9.25	13.80
3.00	7.00	6.00	9.25	8.58	6.25	10.83	5.25	7.60	12.00	8.42	14.67	12.50	11.08	15.58	7.83	11.73
6.00	6.08	4.92	8.33	7.83	5.42	9.00	2.58	6.31	9.00	6.25	10.33	9.83	8.33	11.33	5.75	8.69
Mean B	7.44	6.14	9.36	8.69	6.83	10.78	4.78		11.56	8.92	13.78	12.56	10.69	14.72	7.61	
L.S.D. A				0.4	175							0.5	50			
L.S.D. B				0.0	125							0.8	340			
L.S.D. AxB				1.2	56							1.4	154			-
					2nd san	n ple tak	en at 60 d	l after ti	ansplan.	ting in 2	021					
0.56*	10.08	8.83	10.92	10.67	9.92	11.75	6.67	9.83	13.58	12.08	15.08	12.58	12.92	15.83	8.83	12.99
3.00	8.00	7.08	9.67	9.17	7.50	10.00	5.50	8.13	11.08	9.58	12.67	11.58	10.08	13.75	7.33	10.87
6.00	6.58	4.58	7.92	7.17	5.17	8.58	3.75	6.25	8.33	5.58	10.50	7.92	6.58	10.67	4.63	7.74
Mean B	8.22	6.83	9.50	9.00	7.53	10.11	5.31		11.00	9.08	12.75	10.69	9.86	13.42	6.93	
L.S.D. A				0.4	165							0.6	549			
L.S.D. B				0.0	60,							0.0	16			
L.S.D. AxB				1.2	29							1.7	117			

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\*= tap water (control) \*\*= The same foot notes as indicated in Table 7

0	ld leaves	** of tom	ato plan	ts determ	ined at 50	0d (in 20	20 & 202	1) and 6	0d (in 20	(21 only)	after tra	insplantir	Bi			
							Salinity	r alleviat	ion trea	tments						Q
Salinity						Sample	e taken at	50 d aft	er transj	planting	in 2020					
levels dS/m	Na con	tent in y	oung lea	ves (mg/k	g)			-					Na cont	ent in ol	ld leaves (	mg/kg
(Y)	MA	PGPR	MA+	COM	HA	COM	Untreat.	Mean	MA	PGPR	MA+	COM	HA	COM	Untreat.	Mean
0.56*	9 12.	100cu. 9 94	7 98	AppIIC. 8 29	Applic. 9 12	7 50	control 10.83	A 8 97	21 75	22.05	20.01	20 58	21 81	+ HA 14 43	23 56	20.60
3.00	28.32	31.47	27.02	26.70	28.00	25.75	35.64	28.99	36.35	41.20	33.48	34.55	38.54	30.68	42.75	36.79
6.00	45.92	54.29	41.19	43.53	53.38	38.17	58.61	47.87	53.82	59.49	46.73	52.97	55.97	42.75	61.74	53.35
Mean B	27.79	31.90	25.40	26.18	30.16	23.81	35.02		37.31	40.91	33.41	36.03	38.77	29.28	42.68	
L.S.D. A				1.2	274							[C]	10			
L.S.D. B				1.5	046							2.0	02			
L.S.D. AxB				3.3	171							3.4	-67			
					] <sup>st</sup> Sa.	mple tak	en at 50 d	after trar	nsplantin	g in 202]						
0.56*	13.38	16.21	10.84	11.72	13.14	10.38	20.59	13.75	27.02	29.32	22.36	23.25	25.10	18.60	34.16	25.69
3.00	37.45	40.83	34.16	35.60	38.18	26.06	48.01	37.18	44.72	47.94	42.75	43.93	47.39	25.76	52.53	43.57
6.00	42.75	49.18	37.06	38.99	47.94	35.97	54.24	43.73	57.29	64.54	55.18	56.44	59.05	50.42	71.56	59.21
Mean B	31.20	35.40	27.35	28.77	33.09	24.14	40.95		43.01	47.27	40.10	41.21	43.85	31.59	52.75	
L.S.D. A				1.1	81							2.8	67			
L.S.D. B				1.8	304							4.3	80			
L.S.D. AxB				3.1	.25							7.5	86			
					2nd s	ample tak	cen at 60 c	l after tra	insplantii	10 in 202	1					
0.56*	10.60	14.87	9.31	9.74	12.20	8.09	18.04	11.83	20.90	22.05	18.04	19.15	21.45	16.70	24.49	20.40
3.00	44.73	49.60	40.83	42.75	46.72	35.97	57.73	45.48	55.10	71.11	47.94	51.26	58.62	47.94	82.47	59.21
6.00	72.04	76.44	64.51	65.75	74.02	60.44	88.20	71.63	92.93	100.06	82.30	88.70	96.22	75.97	103.37	91.36
Mean B	42.46	46.97	38.21	39.41	44.31	34.83	54.66		56.31	64.41	49.43	53.04	58.76	46.87	70.11	
L.S.D. A	~			1.5	36							1.2	92			
L.S.D. B				2.3	346							1.5	74			
L.S.D. AxB				4.0	)64							3.4	.19			
*= tan water (	control)															

Table (11): Effect of salinity levels (A) and some salinity alleviation treatments (B) and their interactions (A×B) on sodium (Na) content in young and

\*\*= The same foot notes as indicated in Table 7.

Alleviation salinity stress on tomato plants by some organic and bio-fertilizers application

able (12):	Effect of old leave	salinity s* of tor	levels (A nato pla	<ul> <li>v) and so</li> <li>nts deteri</li> </ul>	me salini mined at	ty allevia 50 d (in 3	tion treat 2020 & 20	tments (] )21) and	8) and th 60 d (in :	ieir inter 2021only	actions ( ) after ti	(A×B) or ransplan	ı chloride ting.	(Cl) con	tent in yo	oung and
							Salinit	y allevia	tion trea	tments						Γ
Salinity						Sampl	e taken ai	t 50 d afi	er trans	planting	in 2020					
dC/m	Cl con	itent in y	oung lea	ives (mg/	kg )								Cl cont	ent in old	leaves (1	ng/kg)
	MA	PGPR	MA+	COM	HA	COM+	Untreat.	Mean	MA	PGPR	MA+	COM	HA	COM+	Untreat.	Mean
(m)	Inocu.	Inocu.	PGPR	Applic.	Applic.	HA	control	А	Inocu.	Inocu.	PGPR	Applic.	Applic.	HA	control	А
0.56*	92.67	100.33	92.33	90.33	99.67	86.00	103.00	95.28	101.67	105.00	98.33	97.67	104.00	91.00	106.33	100.57
3.00	123.67	130.33	120.67	101.33	130.33	89.33	132.33	117.39	132.00	134.67	130.67	117.67	135.00	93.67	138.67	126.05
6.00	133.33	136.67	127.67	118.33	136.00	95.33	139.00	125.50	139.33	141.00	134.67	131.67	140.33	105.00	144.00	133.71
Mean B	116.56	122.44	113.56	103.33	122.00	90.22	124.78		124.33	126.89	121.22	115.67	126.44	96.56	129.67	
L.S.D. A				1.	603							2.	497			
L.S.D. B				5	448								814			
L.S.D. AxB				4	241							9.	606			
					1 <sup>st</sup> 5	sample tal	ken at 50 d	d after tre	unsplantir	12 in 202	1					
0.56*	98.33	104.33	90.00	98.00	101.00	82.33	107.00	97.29	101.67	117.67	92.33	98.67	107.67	91.33	124.67	104.86
3.00	111.33	116.33	96.33	99.67	111.33	92.00	125.33	107.48	121.67	129.67	108.67	111.33	125.67	96.33	131.67	117.86
6.00	125.33	123.67	103.33	118.00	119.67	95.67	127.67	116.19	128.67	130.67	120.67	124.67	129.67	98.33	133.33	123.71
Mean B	111.67	114.78	96.56	105.22	110.67	90.00	120.00	111.67	117.33	126.00	107.22	111.56	121.00	95.33	129.89	
L.S.D. A				1.	469			2				2.	008	20 E		
L.S.D. B				2.	243							3.	056			
L.S.D. AxB				З.	886							5.	294			
					2nd	sample te	aken at 60	d after tr	ansplanti	ng in 202	21					
0.56*	101.33	116.33	91.67	99.00	111.33	87.67	119.00	103.76	109.67	116.67	102.67	103.67	114.00	92.33	122.33	108.76
3.00	130.33	136.00	126.33	129.67	130.67	90.67	138.33	126.00	136.67	141.67	129.33	133.00	137.67	98.33	146.33	131.86
6.00	137.67	137.67	136.67	137.00	139.00	102.67	141.67	133.19	143.33	146.67	141.00	142.00	146.67	109.00	148.67	139.62
Mean B	123.11	130.00	118.22	121.89	127.00	93.67	133.00		129.89	135.00	124.33	126.22	132.78	99.89	139.11	
L.S.D. A				1.	145							1.	347			
L.S.D. B				1.	749							2.	057			
L.S.D. AxB				ς.	029							ς.	563			
	-41-11V															5

\*= tap water (control)

\*\*= The same foot notes as indicated in Table 7.

In general, data in Tables 7 - 10 indicated that salinity alleviation treatments enhanced useful mineral nutrient contents in tomato leaves in both normal (non-saline) and saline conditions, but their effect was more pronounced under normal conditions. Also, such treatments increased N. P. K and Ca contents of tomato leaves in both young and old leaves, this increment was slightly decreased in samples taken at 60 DAT than those determined at 50 DAT. As previously mentioned, that salinity alleviation treatments increased N, P and K contents in young and old leaves but the proportion of the increment in young leaves was higher than that observed in old ones. However, Na and Cl contents in both young and old leaves both were decreased by using salinity alleviation treatment and the depression was more pronounced in young than in old leaves, this may be in line with the well-known knowledge that one mechanism in alleviation salinity hazard of plants is to motivate toxic ions e.g. Na and Cl to accumulate in old nonactive leaves. The accumulation of Na and Cl in older leaves while their concentrations remain low in younger leaves is an important physiological trait and salt tolerant mechanism to reduce salt accumulation in young active leaves (Soliman and Does, 1992 and Cuartero and Fernadez-Munoz, 1999).

All salinity alleviation treatments (either bio or organic fertilizers) enhanced N, P, K, and Ca content in tomato young and old leaves under saline conditions, (Tables, 7, 8, 9 and 10). But N, P and K content was higher in younger leaves than older ones, however, Ca content shows a countertrend as its content in older leaves was higher than in younger ones (Table 10). The higher concentration of calcium in older leaves (at the bottom of plants) compared to that in younger ones (upper leaves) observed in this study be returned turn to the special trait of calcium which is among those elements that move slowly in plants and its upward movement takes place in the transpiration stream (TS) through the xylem, TS fall down as a response to stomata closure caused by salinity, which more restricted Ca upward movement, this may explains the high concentration of Ca in lower old leaves, under the condition of this study.

It was demonstrated that AM inoculation of tomato plants grown under saline condition

improved the uptake of almost essential nutrients (such as nitrogen, potassium, calcium and phosphors) by tomato plants (Balliu et al., 2015) while decrease the uptake and transport of Na+ in pepper plants grown under saline conditions (Cekic et al., 2012), and reducing the uptake of toxic ions such as Na and Cl in wheat plants irrigated with saline water (Daei et al., 2009). Also, similar findings were obtained by Hajiboland et al., (2010) who found that AM inoculation alleviated salt-induced reduction of P, Ca and K uptake in tomato and enhanced Ca/Na and K/Na ratios. PGPR in addition, can increased mineral ions via stimulation of proton pump ATPase (Mantelin and Touraine, 2004). Thus, Karlidage et al. (2013) reported that strawberry plants grown under salinity stress and inoculated with PGPR significantly increased element contents of leaves such as N, K, P and Ca. Moreover, Bacillus subtilis also enhanced nitrogen fixation and solubilize soil P (Hashem et al., 2019).

Enrichment of organic matter in the soil leads to improve soil physical, chemical and biological properties, increased soil dissolved organic C and nutrient retention capacity of salt-effected soil and improving plant nutrient use efficiency (Qadir and Oster 2004, Clark et al, 2007 and Wang et al., 2014). Therefore, COM application resulted in the enhancement of plant nutrient uptake and accumulation in tomatoes (walker and Bernal, 2008), eggplant (Semida et al., 2014) and in barley (Liang et al., 2005 and Tejada et al., 2006) plants grown under saline conditions. Similarly, Leogrande et al. (2016) mentioned that COM application significantly decreased the sodium adsorption rate and increased potassium and calcium contents on tomato plants which were irrigated with saline water (EC=6.0 dS/m).

While, it was also reported that the mechanism of HA in promoting plant growth may be by enhance the uptake of useful nutrients and reduce the uptake of toxic elements such as Na and Cl (Knicker *et al.*, 1993, Tan, 1998 and Friedel and Scheller, 2002). HA application also was able to improve N, P and K contents in tomato plants leaves that were grown in saline conditions (Ashraf and Mohamed 2008 and Feleafel and Mirdad 2014a) while the reverse was true for Na and Cl (Ashraf and Mohamed 2008).

#### **4-2-** Proline content in tomato leaves

Proline is one of the compatible organic solutes that are used by plant as osmoprotectant under stress conditions. The data presented in Table 13 clearly show that proline content in tomato leaves was significantly increased by salinity and its increment was more pronounced with highest salinity level of irrigation water i.e. 6dS/m, in both seasons. Thus, these findings support the previous findings (Azami *et al.*, 2010, Eraslan *et al.*, 2015 and Ali and Rab, 2017) regarding the increase of proline content in tomato leaves by salinity stress. This accumulation of osmolytes especially proline is a common phenomenon in plants under salt stress.

The salinity alleviation treatments, however, decreased proline content in tomato leaves than those in untreated plants (Table 13). But it is observed that treatments that did well in enhancing vegetative growth, RWC, uptake of benefit nutrient elements and reduced toxic ions uptake (previously found in this study) gave lower proline content than those treatments that had less influence on growth parameters and other traits that enhancing salt tolerance. Such result seems reasonable since the favorable changes which induced salinity mitigation in plants by combined treatments i.e, COM+HA and other treatments reduced the required of further accumulation of proline content.

However, the response of proline content to bio-fertilizer treatments under salinity stress is somewhat contradictory, i.e., some studies demonstrated that AM inoculation increased proline contents in tomato plants (Barin et al., 2006 and Hajiboland et al., 2010, Dargiri et al., 2021). On the other hand other studies indicated that AM untreated plants accumulated more proline than those treated (Jahromi et al., 2008 on lettuce, Kaya et al., 2009 on pepper, and Isfahani et al., 2019 and Turan et al., 2021 on tomato plants) all grown under saline conditions. Thus, it could be concluded that bio-fertilizers application can reduce the severity of salt stress and enhance mitigation of salinity, this may resulted in reduce proline accumulation.

Fable (13): Effect of salinity levels (A), some salinity alleviation treatments (B) and their interactions
$(A \times B)$ on proline content in leaves of tomato plants determined at 50 d after transplanting
in both seasons of study.

G 11 14			Sali	nity alleviat	ion treatment	s (B)		
Salinity			Proli	ne content ir	n leaves (µ/g I	Dr.Wt)		
dS/m				Sease	on 2020			
(A)	Mycorrizal	B. subtilis	Mycorrizal +	Compost	Humic Acid	Compost +	Untreated	Moon A
(11)	Inoculation	Inoculation	B. subtilis	Application	Application	Humic Acid	control	Mean A
$0.56^{*}$	122.75	188.97	119.93	100.54	126.45	90.86	212.68	137.46
3.00	353.58	403.25	311.29	268.43	395.79	236.98	502.16	353.07
6.00	398.84	541.17	462.85	437.41	524.60	415.78	893.04	524.81
Mean B	291.72	377.80	298.03	268.79	348.95	247.87	535.96	
L.S.D A				18	.913			
L.S.D B			28.891 50.040 Senson 2021					
L.S.D AxB								
				Season 2021	l			
0.56 *	103.50	119.20	80.11	90.59	115.58	77.20	138.62	103.54
3.00	238.59	451.96	197.97	217.04	249.64	157.86	442.30	279.34
6.00	451.64	571.87	326.61	337.64	549.58	238.90	746.35	460.37
Mean B	264.58	381.01	201.57	215.09	304.93	157.99	442.43	
L.S.D A				26	.673			
L.S.D B				40	.744			
L.S.D A x B				70	.571			

\*= tap water (control)

Proline content in plant leaves also shows a contradictory response toward the effect of organic fertilizer under salinity conditions (Table 13). El-Galad et al. (2013) in a similar work found that compost treatment of faba bean plants grown under saline conditions significantly decreased proline content. Hammad et al. (2010) explained that organic fertilizer maintains osmotic adjustment to keep continuous water absorption at low soil water potential caused by salinity, such favorable effect of organic fertilizer reduced salinity detrimental effect on plants so that decrease plants requirement of proline. On the other hand Rady (2012) on tomato and Semida et al. (2014) on eggplant both grown under saline conditions, showed an increase in proline contents in plants fertilized with organic fertilizer compared to those of untreated control plants.

The interaction effect between salinity levels and salinity alleviation treatments on proline content (Table 13) shows that the significantly highest value of proline content was obtained by untreated control with 6 dS/m in both seasons. However, the lowest value of proline was recorded to the combined treatment of COM+HA with 0.56 dS/m level of salinity (Table 13). Proline in plants treated with salinity alleviation treatments and irrigated with tap water (0.56 dS/m) show lower values than that obtained from counterpart treatments but subjected to salinity levels i.e.,(3.0 and 6.0dS/m).

#### 5- Electrolyte leakage (EL)

According to the data given in Table 14 electrolyte leakage (EL) tended to increase consistently and significantly with each increase in salinity level in irrigation water of tomato, and the proportion of the increment aggravated at 6 dS/m than at 3 dS/m of salinity level. These results seemed to be accordance with those obtained by (Manaa *et al.*, 2011, Tartoura *et al.*, 2014 and Ors *et al.*, 2021) who reported that EL values increased proportionally in tomato leaves with increasing salt concentration.

Table (14): Effect of salinity	levels (A), some salinity alleviation treatments (B) and their interactions
(A×B) on electro	lyte leakage in tomato leaf determined at 50 d after transplanting in both
seasons of study	

			Salinity al	leviation tro	eatments (B)	)		
Salinity			Electroly	yte leakage	values (%)			
levels				Season 202	20			
dS/m (A)	Mycorrizal Inoculation	B. subtilis Inoculation	Mycorrizal + B. subtilis	Compost Application	Humic Acid Application	Compost + Humic Acid	Untreated Control	Mean A
0.56*	47.43	57.58	46.78	45.96	54.80	32.84	61.54	49.56
3.00	73.84	81.41	67.99	66.38	76.66	66.14	87.33	74.25
6.00	83.84	82.70	78.45	73.39	85.95	69.62	90.64	80.66
Mean B	67.87	73.54	62.36	66.39	72.47	56.20	79.84	
L.S.D A				0.683				
L.S.D B				1.044				
L.S.D AxB				1.808				
			Seaso	n 2021				
0.56 *	31.04	43.25	27.63	28.25	36.70	26.47	49.57	34.70
3.00	58.33	63.75	50.24	55.55	60.81	45.63	70.91	57.81
6.00	73.00	77.97	69.52	72.70	75.93	66.32	87.08	74.64
Mean B	54.12	61.65	49.13	51.99	57.81	46.14	69.19	
L.S.D A				1.318				
L.S.D B				2.013				
L.S.D A x B				3.486				

\*= tap water (control)

According to Manaa *et al.* (2011) EL is known as an indicator of membrane damage caused by salt stress in tomato leaves according to NaCl concentration. Also, salinity induces reactive oxygen species (ROS) formation which can lead to oxidative damage in various cellular components such as proteins and lipids particularly those in cell membrane (Apel and Hirt, 2004, Munns and Tester, 2008, Rahnama *et al.*, 2010 and Ahmed and Umar, 2011). These findings may explained why EL is associated with stress conditions particularly salinity.

Salinity alleviation treatments, on the other hand seriously mitigate the hazard effect of salinity so that they all decreased EL. Also, treatments that did well in enhancing growth, useful nutrient uptake and improve water status, previously mentioned in this study (i.e., combined treatments of COM+HA, and AM+PGPR and AM and COM applied alone) gave the lower values of EL (Table 14). These results support the former reports regarding the favorable effect of AM in reducing EL in cucumber plants (Ahmad et al., 2019) and in pepper plants (Kaya et al., 2009), both grown in saline conditions. Also, similar findings were mentioned by Bano and Fatima (2009) on Zea Maize, Karlidag et al. (2013) on strawberry and Ullah et al. (2016) on tomato, who observed that PGPR decreased EL in cells of plants suffer from salinity stress. Among the roles of biofertilizers (particularly AM) as salinity alleviation treatment used in this study is to enhance the synthesis of antioxidant enzymes (Aguilar- Aguilar et al., 2009) and also increase their activity (Heikham et al., 2009) for scavenging of ROS. Also, Rady et al. (2016) found that application of organo-mineral fertilizer compost significantly reduced EL in bean plants which grown in saline soil. HA, as well, added to saline soil significantly reduced EL in bean plants (Aydin et al., 2012). In addition, compost as organic fertilizer has the capability to increase antioxidants activities which enhance salt tolerance to salinity and other stress conditions. Moreover, salinity alleviation treatments used in this study reduced toxic elements i.e., Na and Cl uptake, and enhance water content and nutrient element uptake such favorable conditions would

reduce salinity detrimental effect on cell membrane and reduce EL.

The highest value of electrolyte leakage of tomato plants (Table 14) was obtained by the untreated control with 6 dS/m level of salinity i.e.90.64 and 87.08 % in 2020 and 2021 seasons respectively, however the lowest values were 32.84 and 26.47 in the same order were recorded to the combined treatment of COM+HA with 0.56 dS/m (Table 14).

As expected the combined treatment of COM+HA with 0.56 dS/m level gave significantly the lowest value of electrolyte leakage in tomato leaves, in both seasons.

#### 6- Fruit weight and total yield

#### 6-1- Average fruit weight

Results obtained in Table 15 indicate that salinity reduced average fruit weight, and the reduction tended to decrease consistently and significantly with each increase in salinity level in both seasons. Accordingly, the reduction percentages (average of the two seasons) than that fruit weight of non-saline treatment were 17.9 and 29.0 % at 3 and 6 dS/m respectively.

These results seemed to be in accordance with those obtained by Greenway and Munns (1980), Magan et al. (2008) and Zhai et al. (2016) regarding the detrimental effect of salinity on average fruit weight of tomato. It was also previously mentioned that the reduction in average tomato fruit weight occurred even at low and moderate salinity levels; i.e., at 3-4 dS/m (Malash et al., 2008, Scholberg and Locascio 1999), but the reduction was more pronounced at higher salinity level i.e. 9.6 dS/m<sup>-1</sup> (Souza, 1990 and Al-Yahyai et al., 2010). Such reduction in average fruit weight by salinity could be explained by the fact that salinity particularly high levels decreased water potential of tomato plants which reduces water flow into fruit and limit the rate of fruit expansion (Johnson et al., 1992 and Al-Ismaily et al., 2014). Also, the accumulation of Na in tomato plant leads to such reduction in mean fruit weight of tomato (Adams, 1991 and Cuartero and Fernandez-Munoz, 1999).

			Salini	ty alleviation	treatments (	<b>B</b> )	Untreated control M 23.91 2 20.20 2 17.15 2 20.42 15.73 1 12.23 1 8.90 1 15.73	
Salinity			A	verage fruit	weight (g)			
levels				Season	2020			
dS/m (A)	Mycorrizal Inoculation	B. subtilis Inoculation	Mycorrizal + B. subtilis	Compost Application	Humic Acid Application	Compost + Humic Acid	Untreated control	Mean A
$0.56^{*}$	28.16	28.23	28.33	27.44	27.59	28.02	23.91	27.38
3.00	23.23	22.47	23.52	23.80	21.72	25.95	20.20	22.99
6.00	20.67	20.68	20.80	21.71	21.56	19.06	17.15	20.23
Mean B	24.02	23.79	24.22	24.32	23.62	24.34	20.42	
L.S.D A				0.40	9			
L.S.D B				0.62	4			
L.S.D AxB				1.08	1			
			Se	eason 2021				
$0.56^{*}$	17.56	16.51	17.63	17.84	16.70	20.10	15.73	17.44
3.00	13.38	13.92	14.11	13.50	13.38	15.38	12.23	13.70
6.00	10.82	10.07	13.82	12.48	10.92	13.25	8.90	11.47
Mean B	17.56	16.51	17.63	17.84	16.70	20.10	15.73	
L.S.D A				0.61	3			
L.S.D B				0.93	7			
L.S.D A x B				1.62	3			

Table (15): Effect of salinity levels (A), some salinity alleviation treatments (B) and their interactions (A×B) on average fruit weight (g) of tomato in both seasons of study.

\*= tap water (control)

Salinity alleviation treatments significantly increased average fruit weight of tomato of both plants grown under non-saline (0.56 dS/m) and saline (3 and 6 dS/m) conditions than those obtained from plants untreated in both seasons of study (Table 15). But differences between treatments were not significant in most cases in 2020 season; i.e., the highest fruit weight was obtained by the combined treatment of COM+HA which was significantly differ only with that obtained by HA treatment applied alone (Table 15), whereas in 2021 the combined treatment of COM+HA gave significantly the highest fruit weight compared to other treatments. The enhancement of salinity alleviation treatments of average tomato fruit weight was also mentioned; i.e., Barin et al., (2006) and Hadad et al., (2012) with AM inoculation, Saha et al. (2017) with compost application and Kumar et al. (2017) with HA application. The favorable effect of such treatments on average fruit weight is expected

since these treatments resulted in improving water status in tomato plants (Table 5) and reduced toxic ions (Na and Cl) uptake (Tables 11and 12).

Regarding the interaction between salinity and alleviation treatments Table 15 shows that the highest value of average fruit weight of tomato plants was recorded to the combination between 0.56 dS/m level of salinity and the combined treatments of AM + *B. subtilis*, and COM+HA in 2020 and 2021 seasons respectively. On the other hand, the lowest values were obtained by the combination between 6.0 dS/m and untreated control in both seasons.

#### 6-2- Total fruit yield of tomato /plant

Table 16 shows that total yield of tomato/ plant decreased consistently and significantly with each increase in salinity level. The reduction percentage (average of the 2 seasons) in total yield were 26.9% at 3 dS/m and 46.8% at 6dS/m<sup>-1</sup>, this implies that each 1dS/m increase in salinity level decreased tomato total yield by 6.6% (among this range of salinity under conditions of this study). These results are in agreement with those reported by Mohammad et al. (1998), Malash et al. (2008), Viol et al. (2017) and Pengfei et al. (2019), regarding the reduction of tomato total yield by exposing to salinity in its root zone. Also, Cuartero and Fernandez-Munoz (1999), Del Amor et al. (2001) and Malash et al. (2008) indicated that tomato yield is quite sensitive to salinity at 3.0 dS/m and above. Moreover, Moghaddam et al. (2018) showed that salinity at 4 dS/m and 7dS/m decreased tomato fruit yield by 27.2% and 46.7% respectively (compared to those without salt stress) which are somewhat similar to the corresponding values of this recent study at 3 and 6 dS/m respectively. Also, Zhang et al. (2016) reported that the reduction rate in fruit yield of tomato with increasing EC unit of salinity equal and above 5dS/m was 7.2%, thus this finding is somewhat similar to corresponding values obtained in this study (mentioned above).

The reason of reducing tomato yield by salinity, may return to higher osmotic pressure in plants (Ayers, 1977, Cuartero and Fernandez-Muroz, 1999 and Zhang *et al.*, 2016), or to the reduction in WUE (Al-Harbi *et al.*, 2009 and Al-Omran *et al.*, 2012) and to accumulation of toxic ions such as Na and Cl (Niu *et al.*, (1995).

Regarding salinity alleviation treatments, Table 16 shows that all treatments increased tomato total yield under saline and non-saline conditions. It is worth mentione hat the effect of combined treatment of COM+HA, as this treatment in particular gave the best performance in alleviation salinity hazard in this study, such treatment increased total yield under saline conditions rather than under normal (non-saline) conditions. This finding was similar to those obtained by Al-Karaki (2006) who indicated that AM inoculated tomato plants showed an enhancement in fruit yield by 24% under nonsaline and 60% under saline conditions.

Table (16): Effect of salinity levels (A), some salinity alleviation treatments (B) and their interactions(A×B) on tomato total yield in both seasons of study.

			Salinity	alleviation tr	eatments (1	<b>B</b> )		
Salinity			Т	otal yield (g/	plant)			
levels				Season 20	20			
dS/m (A)	Mycorrizal Inoculation	B. subtilis Inoculation	Mycorrizal + B. subtilis	Compost Application	Humic Acid Application	Compost + Humic Acid	Untreated control	Mean A
$0.56^{*}$	137.58	123.83	143.17	143.78	128.55	152.23	91.50	131.52
3.00	96.68	85.76	104.27	108.29	86.31	134.27	70.59	98.02
6.00	73.36	68.94	81.95	88.92	75.34	92.39	42.77	74.81
Mean B	102.54	92.84	109.80	113.66	96.73	126.30	68.29	
L.S.D A				0.423				
L.S.D B				0.737				
L.S.D AxB				1.277				
			Seas	on 2021				
$0.56^{*}$	112.10	79.38	126.05	118.03	98.64	151.80	71.86	108.27
3.00	77.82	64.77	92.57	83.07	74.52	102.39	44.62	77.11
6.00	48.44	37.36	73.10	58.30	45.56	77.10	26.89	52.39
Mean B	79.45	60.50	97.24	86.46	72.91	110.43	47.79	
L.S.D A				0.931				
L.S.D B				1.421				
L.S.D A x B				2.462				

\*= tap water (control)

The enhancement of total fruit yield of tomato grown under saline condition by AM was also mentioned elsewhere (Barin et al., 2006, El-Shazly, Abdelhameid and 2020 and Pietrantonio et al., 2020). The beneficial effect of AM on yield of tomato grown in saline conditions were: provides nutrition to the host plants, as well as increasing water uptake, production of hormones and enhancing adaptation to environmental stress including salinity (Garg and Chandel, 2010). The improvement of total fruit yield grown in saline stress induced by PGPR was also mentioned elsewhere (Aini et al., 2021, Turan et al., 2021 on tomato and Bochow et al., 2001 on eggplant and pepper). PGPR fixing atmospheric nitrogen, producing phytohormones, solubilizing minerals (Mayak et al., 2004a), enhancing reactive oxygen species (ROS) scavenging (Jianmin et al., 2014) reduce salt toxicity by lowering the Na concentration in plants (Abd El-Samad et al., 2004, Yildirim et al., 2006 and Kohler et al., 2009), and reduces synthesis of harmful ethylene (Glick, 2014). This favorable effect of each treatment applied alone on total yield will be aggravated when both (AM+PGPR) added together which gave a synergistic effect observed in this study.

Compost (COM) application also enhanced tomato fruit yield even grown under saline conditions (Rady,2012 and Saha *et al.*, 2017). Also, the favorable effect of HA application on fruit yield of tomato plants grown under saline stress was also previously reported (Feleafel and Mirdad, 2014a, feleafel and Mirdad, 2014b and Kumar *et al.*, 2017). The benefit obtained by COM application to plants grown under saline conditions was improving soil physical, chemical and biological properties (Qadir and Oster, 2004, Walker and Bernal, 2008, and Wang *et al.*, 2014), such conditions enriched soil by humic substances, macro and micro-nutrients (Walker and Bernal, 2008 and Wright *et al.*, 2008).

The useful advantages of HA application in mitigation salinity hazard which dramatically reduces yield of plants were: enhancing the uptake of beneficial nutrient elements and reduce the uptake of toxic elements (Knicker *et al.*, 1993, Tan, 1998 and Friedel and Scheller, 2002),

transportation and availability of micro nutrient (Bohme and Lua, 1997) and by changing ion balance and enhancing photosynthesis rate (Souza *et al.*, 2021).

Gathering the above mention advantage of COM and HA in one treatment of course will give a synergistic effect that was showed in the recent study.

According to the data of the interaction between salinity levels and salinity alleviation treatments, Table 16 shows that the highest total yield of tomato/ plant obtained was a result of using the combined treatment of COM+HA along with 0.56 dS/m salinity level (un-saline). The 2nd highest value of tomato total yield/plant was recorded to the COM treatment in 2020 season, but such rank was recorded to the combined treatment of AM+PGPR in 2021 season, both along with 0.56 dS/m salinity level. While the lowest values of tomato total yield/plant were due to those plants subjected to the highest salinity level (6.0 dS/m) and untreated with any of these salinity alleviation treatments (Table 16). HA application and PGPR inoculation along with the highest salinity level gave also lower total fruit yield /plant (Table 16).

### 7- Fruit quality

# 7-1 Total soluble solids (TSS) content in tomato fruit

Salinity increased TSS content in tomato fruits and the increase was growing with increasing salinity levels in the irrigation water in both seasons (Table 17). This result agreed with former reports regarding the positive effect of salinity on tomato fruit quality including TSS (Mizrahi et al., 1988, De Pascale et al., 2001, Malash et al., 2002 and Maggio et al., 2004). Table 17 also shows that TSS values were higher in the two salinity levels than those obtained by non-saline treatment whatever was salinity alleviation treatments used. The reason of the increase in TSS content in tomato fruit by salinity was clarified by several researchers, such reasons are:1- salinity promotes starch accumulation in immature tomato fruit which consider as a reservoir for soluble sugars accumulation during fruit ripening ,contributing

to the final fruit sugar level (Schaffer *et al.*, 2000 and Petreikav *et al.*, 2009), 2- the increase of tomato fruit soluble solids seems to be associated with the reduction in the water content of the fruit, (Adams and Ho 1989,Cuartero and Fernandez-Munoz,1999 and Magan *et al.*, 2008), and 3- the increasing in total soluble solids by salinity may due to smaller fruit size (Ho *et al.*, 1996).

Tomato plants that subjected to salinity alleviation treatments, however produced fruits with lower TSS content than those produced by untreated plants, but differences were not significant in most cases (Table 17). Such treatments which resulted in reducing TSS may mitigated salinity effect in a way that enhanced water status (Table 5), and increase fruit size and weight (Table 15) such conditions reduced TSS in fruits. The reduction in TSS contents in tomato fruits by salinity alleviation treatments was also observed by Al-karaki and Hammad (2001) who mentioned that TSS content in tomato fruits of plants inoculated with AM was lower than those obtained from plants un-inoculated when both plants grown in saline condition. The same authors added that stress conditions induced by salinity enhances fruit quality of tomato, while AM treatment mitigate the harmful effect of salinity by improve water and nutrient status as well as another physiological and biochemical process, such favorable effect reduced TSS content.

Also, the non-significant differences in TSS content of tomato plants inoculated by AM and those of uninoculated plants both grown under saline conditions were also recorded by Huang *et al.* (2013). On the other hand Ebrahim and Saleen (2017) and Al-Karaki (2006) indicated that TSS in tomato fruits was higher in AM treated plants than those of untreated plants either grown under saline and non-saline conditions.

Table (17): Effect of salinity levels (A), some salinity alleviation treatments(B) and their interaction
(A×B) on TSS content in tomato fruits determined in mature red fruits one time during
harvesting period in both seasons of study.

			Salinit	y alleviation	treatments	s (B)		
Salinity				TSS conte	ent (%)			
levels				Season	2020			
dS/m (A)	Mycorrizal Inoculation	B. subtilis Inoculation	Mycorrizal + B. subtilis	Compost Application	Humic Acid Application	Compost + Humic Acid	Untreated control	Mean A
$0.56^{*}$	6.68	6.37	6.67	6.75	6.62	6.62	6.25	6.56
3.00	7.55	7.65	7.58	7.60	7.13	6.93	7.98	7.49
6.00	8.07	8.23	8.32	8.17	8.05	8.18	8.68	8.24
Mean B	7.43	7.42	7.52	7.51	7.27	7.24	7.64	
L.S.D A				0.16	9			
L.S.D B				0.25	9			
L.S.D AxB				0.44	8			
			Sea	ason 2021				
$0.56^{*}$	3.68	4.33	3.73	4.40	4.78	4.83	3.95	4.24
3.00	7.08	6.30	6.60	6.78	7.28	7.10	7.68	6.97
6.00	8.33	7.10	7.50	7.53	7.93	8.18	8.45	7.86
Mean B	6.36	5.91	5.94	6.23	6.66	6.70	6.69	
L.S.D A				0.26	6			
L.S.D B				0.40	7			
L.S.D A x B				0.70	5			

\*= tap water (control)

PGPR treatment in this study resulted in produce fruits with TSS values either were not significantly differ (in 1<sup>st</sup> season) or significantly less (in 2<sup>nd</sup> season) than those produced by plants of control (untreated). Thus, these results is not in agreement with those of Shen *et al.*, (2012) who suggested that PGPR was able to improve total and water dissolved sugars under saline conditions.

Table 17 shows also that applied both COM and HA decreased significantly TSS content of tomato fruits (in 2020 season) but such treatments resulted in obtaining fruits had TSS values were not significantly different (in 2021 season) than those produced from the untreated plants and grown under saline conditions.

In previous reports, HA effect on TSS of tomato fruit was also differ, i.e. Ashraf and Mohamed (2008) found significant increase in TSS content of tomato fruit with HA treatment under saline conditions, however Casiorra-Posada and Fischer (2009) found that HA application to tomato plants grown under saline conditions reduced total solids in fruits.

The highest value of TSS of tomato plants (Table 17) was obtained by the untreated control at 6 dS/m level of salinity i.e. 8.68 and 8.45 % in 2020 and 2021seasons respectively, however the lowest values in the same order were recorded to the untreated control in 2020 and mycorrhizal inoculation in 2021 season both at 0.56 dS/m<sup>-1</sup> (Table 17).

# 7-2 Vitamin C (Vit C) content in tomato fruits

Results in Table 18 show that vit C content in tomato fruit increased by salinity and the increase was consistently and significantly with each increase in salinity level, in both seasons. These results support the former reports regarding the enhancement effect of salinity on Vit.C content in tomato fruits (Eraslan et al., 2015, Zhai et al., 2015, Helaly et al., 2017 and Rani et al., 2017). The increase in vit. C content in tomato fruits under salinity stress may be a consequence of the accumulation of monosaccharides in fruits (Cuartero and Fernandez-Munoz, 1999) such monosaccharides were previously mentioned before in TSS discussion. By the way, the chemical symbol of Vit. C is (C<sub>6</sub>H<sub>8</sub>O<sub>6</sub>) which is quite similar to those of monosaccharides (C<sub>6</sub>H<sub>12</sub>O<sub>6</sub>). In addition, the reduction in plant foliage growth by salinity, may increase the exposure of fruits to sunlight which is effective in increasing Vit. C (Radwan et al., 1979 and Malash et al., 2002).

Salinity alleviation treatments increased Vit.C content in fruit of tomato plants than those untreated, in both seasons {with one expetion. i.e. the Vit. C value in fruits produced from plants treated with PGPR was not significantly different than those of plants untreated (control) in both seasons}. The enhancement of Vit.C in tomato fruit by salinity alleviation treatments used in this study (under saline conditions) was also observed by Shen et al. (2012) who mentioned that, from three PGPR strains studied, WP8 strain had the most significant effect in improving Vit.C in fruits of tomato plants grown under saline conditions. Also, Oztekin et al. (2013) found that inoculated tomato plants with AM increased the vitamin C in fruits when plants grown under salinity conditions.

Using organic fertilizers such as COM and HA also enhanced Vit.C content in fruit of tomato plants treated with amended saline irrigation water with humic acid than those obtained without HA application (Ashraf and Mohamed, 2008).

Fable (18):	Effect of salinity levels (A), some salinity alleviation treatments (B) and their interactions
	(A×B) on Vit.C content in tomato fruits determined in mature red fruits one time during
	harvesting period in both seasons of study.

G - 1' '4	Salinity alleviation treatments (B)									
Salinity levels dS/m	Vit. C. content(mg/100g f w)									
	Season 2020									
( <b>A</b> )	Mycorrizal Inoculation	B. subtilis Inoculation	Mycorrizal + B. subtilis	Compost Application	Humic Acid Application	Compost + Humic Acid	Untreated control	Mean A		
0.56*	21.33	19.87	22.40	23.07	20.00	23.20	19.73	21.37		
3.00	24.79	20.02	25.13	27.30	22.62	30.33	19.41	24.23		
6.00	30.77	25.65	30.85	29.73	29.21	35.36	29.29	30.12		
Mean B	25.63	21.85	26.13	26.70	23.94	29.63	22.81			
L.S.D A	0.755									
L.S.D B	1.154									
L.S.D AxB	1.998									
Season 2021										
0.56*	20.72	17.29	21.30	22	19.71	23.36	16.17	20.08		
3.00	26.58	22.18	27.63	28.69	25.87	27.98	20.42	25.62		
6.00	31.50	26.22	34.32	36.43	30.80	37.49	25.70	31.78		
Mean B	26.27	21.90	27.75	29.04	25.46	29.61	20.76			
L.S.D A	1.305									
L.S.D B	1.994									
L.S.D A x B	3.454									

\*= tap water (control)

#### REFERENCES

- Abdelhameid, N.M. and El-Shazly, M.M. (2020).
  The impact of inoculation with arbuscular mycorrhizal fungi on tomato tolerance to salt stress and nutrients uptake in sandy soil. J. of Agri. Chem. and Biotech. Mansoura Univ. 11: 63 70.
- Abd El-Samad Hamdia, M.; Shaddad, M.A.K. and Doaa, M.M. (2004). Mechanism of salt tolerance and interactive effect of Azospirillum bran- silense inoculation on maize cultivars grown under salt stress conditions. Plant Growth Regul. 44: 165–174.
- Adams, P. and Ho, L.C. (1989). Effect of constant and fluctuating salinity on the yield, quality and calcium status of tomatoes. J. Hort. Sci. 64: 725-732.

- Adams, P. and Ho, L.C. (1992). The susceptibility of modern tomato cultivars to blossom end rot in relation to salinity. J. Hort. Sci. 67: 827 -839.
- Adams, P. (1991). Effects of increasing the salinity of the nutrient solution with major nutrients or sodium chloride on the yield, quality and composition of tomatoes grown in rockwool. J. Hort. Sci. 66:201–207.
- Adani, F.; Genevi, P.; Zaccheo, P. and Zocchi, G. (1998). The effect of commercial humic acid on tomato plant growth and mineral nutrition. J.of Plant Nut. 21: 561-575.
- Aguilar-Aguilar, S.; Perez-Moreno, J.; Ferrera-Cerrato, R.; Grimaldo-Juarez, O.; Cervantes-Diaz, L. and Gonzalez-Mondoza, D. (2009). Ectomycorrhiza fungi and tolerance to salinity

in plants. Revista Chilena de Historia Natural.82: 163.

- Ahmad, H.; Hayat, S.; Ali, M.; Liu, H.; Chen, X.; Li, J. and Cheng, Z. (2019). The protective role of 28-homobrassinolide and *Glomus versiforme* in cucumber to withstand saline stress. Plants (Basel). 42:1-18.
- Ahmed, P. and Umar, S. (2011). Oxidative stress: rol of antioxidants in plants.Studium press. New Delhi.
- Aini, N.; Yamika, W.S.D.; Aini, L.Q. and Kurniawan, A.P. (2021). Application of saline tolerant bacteria and soil ameliorants improved growth, yield and nutrient uptake of tomato in saline land. A.J.C.S. 15: 827-834.
- Al-Harbi, A.R.; Wahb-Allah, M.A and Al-Omran, A.M. (2009). Effects of salinity and irrigation management on growth and yield of tomato grown under greenhouse conditions. Acta Hort. 807: 201-206.
- Ali, S.G. and Rab, A.A. (2017). The influence of salinity and drought stress on sodium, potassium and proline content of *Solanum lycopersicum L*. cv. Rio Grande. Pakistan J. of Bot. 49: 1-9.
- Al-Ismaily, S.S.; Al-Yahyai, R.A. and Al-Rawahy, S.A. (2014). Mixed fertilizer can improve fruit yield and quality of field-grown tomatoes irrigated with saline water. J. Plant Nut. 37: 1981–1996.
- AL-Kahtani, M. D. F.; Attia, K. A.; Hafez, Y. M.; Khan, N.; Eid, A. M.; Ali, M. A. M. and Abdelaal, K. A. A. (2020). Chlorophyll fluorescence parameters and antioxidant defense system can display salt tolerance of salt acclimated sweet pepper plants treated with chitosan and plant growth promoting rhizobacteria. Agronomy. 10, 1180:1-20.
- Al-Karaki, G.N., (2006). Nursery inoculation of tomato with arbuscular mycorrhizal fungi and subsequent performance under irrigation with saline water. Sci. Hort. 109:1-7.
- Al-Karaki, G.N. and Hammad, R. (2001). Mycorrhizal influence on fruit yield and mineral content of tomato grown under salt stress. J. of Plant Nut. 24: 1311-1323.

- Al-Omran, A.M.; Al-Harbi, A.R.; Wahb-Allah, M.A.; Alwabel, M.A.; Nadeem, M. and Eleter, A. (2012). Management of irrigation water salinity in greenhouse tomato production under calcareous sandy soil and drip irrigation. J. Agric. Sci. Technol. 14: 939-950.
- Altome, M.M.M.; Maher, G.N.; Magda, A.H. and Abou El seoud, I. I. (2015). Response of tomato plant to compost application and inoculation with mycorrhizal fungi under salt stress conditions. J. Adv. Agric. Res. 20: 46-65. (Fac. Agric. Saba Basha).
- Altunlu, H. (2020). The effects of mycorrhiza and rhizobacteria application on growth and some physiological parameters of pepper (Capsicum annuum L.) under salt stres. Ege Universitesi Ziraat Fakultesi Dergisi, 57(4): 501-510.
- Al-Yahyai, R.; Al-Ismaily, S. and Al-Rawahy, S. A. (2010). Growing tomatoes under saline field conditions and the role of fertilizers. College of Agricultural and Marine Sciences Sultan Qaboos University Sultanate of Oman, pp. 83-88.
- AOAC (1995). Official method s of Analysis of the Association of official Analytical Chemists. Published by the A.O.A.C. International 16<sup>th</sup> ed. Washington, D.C.
- Apel, K. and Hirt, H. (2004). Reactive Oxygen Species: Metabolism, Oxidative Stress, and Signal Transduction. Annual Rev. of Plant Biol. 55: 373-379.
- Arancon, N.Q.; Edwards C.A.; Bierman P.; Metzger J.D.; Lee S. and Welch C. (2003). Effects of vermicomposts on growth and marketable fruits of field-grown tomatoes, peppers and strawberries. Pedobiologia, 47: 731–735.
- Ashraf, S.O. and Mohamed, S.A.E. (2008). The possible use of humic acid incorporated with drip irrigation system to alleviate the harmful effects of saline water on tomato plants. Fayoum J. Agric. Res. & Dev., 22: 52-70.
- Aydin, A.; Kant, C. and Turan, M. (2012). Humic acid application alleviates salinity stress of bean (*Phaseolus vulgaris L.*) plants decreasing membrane leakage. African J. of Agric. Res. 7: 1073-1086.

- Ayers, R.S. (1977). Quality of water for irrigation. Journal of Irrigation and Drainage Division. 103: 135-154.
- Azarmi, R.; Taleshmikail, R. D. and Gikloo, A. (2010). Effects of salinity on morphological and physiological changes and yield of tomato in hydroponics system. J. of Food, Agric. & Environ. 8: 573-576.
- Balliu, A.; Sallaku, G. and Rewald, B. (2015). AMF inoculation enhances growth and improves the nutrient uptake rates of transplanted, salt-stressed tomato seedlings. Sustainability. 7: 15967-15981.
- Bano, A. and Fatima, M. (2009). Salt tolerance in Zea mays (L). following inoculation with Rhizobium and Pseudomonas. Biol. Fert. Soils. 45: 405–413.
- Baradar, A.; Saberi-Riseh, R.; Sedaghati, E. and Akhgar, A. (2015). Effect of some bacteria and iron chelators on potato colonization by arbuscular mycorrhiza fungi inoculated by Rhizoctonia. Indian. J. Sci. Technol. 8: 1–5.
- Barin, M.; Aliasgharzade, N. and Samadi, A. (2006). Effects of NaCl-induced and salts mixture salinity on leaf proline and growth of tomato in symbiosis with arbuscular mycorrhizal fungi. Iranian J. of Agric. Sci. 37: 139-147.
- Barrs, H.D. and Weatherly, P.E. (1962). A reexamination of the relative turgidity technique for estimating water deficit in leaves. Aust. J. Biol. Sci. 15: 413-428.
- Basak, H.; Demdr, K.; Kasim, R. and Okay, F.Y. (2011). The effect of endo-mycorrhiza (VAM) treatment on growth of tomato seedling grown under saline conditions. Afric. J. of Agric. Res. 6: 2532-2538.
- Bates, L.S.; Waldren, R.P. and Teare, I.D. (1973). Rapid determination of free proline for water stress studies. Plant and Soil, 39: 205-207.
- Besford, R.T. and Maw, G.A. (1975). Effect of potassium nutrition on tomato plant growth and fruit development. Plant Soil, 42: 395– 412.
- Bochow, H.; El-Sayed, S. F.; Junge, H.; Stavropoulou, A. and Schmiedeknecht, G.

(2001). Use of *Bacillus subtilis* as biocontrol agent. IV. Salt-stress tolerance induction by *Bacillus subtilis* FZB24 seed treatment in tropical vegetable field crops, and its mode of action. J. plant Diseases and Protection. 108(1): 21-30.

- Böhme, M. and Lua, H, T. (1997). Influence of mineral and organic treatments in the rizosphere on the growth of tomato plants. Acta Hort., 450: 161-168.
- Boyer, J.S. (1969). Measurements of the water status of plant. Ann. Rev. Plant physiol. 20: 351-364.
- Calvo-Polanco, M.; Sánchez-Castro, I.; Cantos, M.; García, J.L.; Azcón, R.; Ruiz-Lozano, J.M.; Beuzón, C.R. and Aroca, R. (2016). Exploring the use of recombinant inbred lines in combination with beneficial microbial inoculants (AM fungus and PGPR) to improve drought stress tolerance in tomato. Environ. Exp. Bot. 131: 47–57.
- Canellas, L.P.; Olivares, F.L.; Aguiar, N.O.; Jones, D.L.; Nebbioso, A.; Mazzei, P. and Piccolo, A. (2015). Humic and fulvic acids as biostimulants in horticulture. Sci. Hort. 196: 15–27.
- Casierra-Posada, F. and Fischer, G. (2009). Reducing negative effects of salinity in tomato (Solanum lycopersicum L.) plants by adding leonardite to soil. Acta Hort. 821: 133-137.
- Cekic, F. O.; Unyayar, S. and Ortas, I. (2012). Effects of arbuscular mycorrhizal inoculation on biochemical parameters in Capsicum annuum grown under long term salt stress. Turk. J. Bot. 36: 63–72.
- Chapman, H.D. and Pratt, D.F. (1961). Method of analysis for soil, plant and water. Div. of Agric. sci. Univ. of calif.
- Cimrin, K. M.; Turkmen, O.; Turan, M. and Tuncer, B. (2010). Phosphorus and humic acid application alleviate salinity stress of pepper seedlings. African J. Biotech. 9: 5845–5851.
- Clark, G.J.; Dodgshun, N.; Sale, P.W.G. and Tang, C. (2007). Changes in chemical and biological properties of a sodic clay subsoil with addition of organic amendments. Soil Biol. Biochem. 39: 2806–2817.

Alleviation salinity stress on tomato plants by some organic and bio-fertilizers application

- Canellas, L.P.; Olivares, F.L.; Aguiar, N.O.; Jones, D.L.; Nebbioso, A.; Mazzei, P. and Piccolo, A. (2015). Humic and fulvic acids as biostimulants in horticulture. Sci. Hort. 196: 15–27.
- Cruz, V.; Cuartero, J.; Bolarin, M.C. and Romero, M. (1990). Evaluation of characters for ascertaining salt stress responses in *Lycopersicon species*. J. Am. Soc. Hort. Sci. 115: 1000-1003.
- Cuartero, J. and Fernandez-Munoz, R. (1999). Tomato and salinity. Sci. Hort. 78: 83-125.
- Daei, G.; Ardekani, M. R.; Rejali, F. and Teimuri, S. (2009). Alleviation of salinity stress on wheat yield, yield components and nutrient uptake during arbuscular mycorrhizal fungi under field conditions. J. Plant Physiol. 166: 617–625.
- Dargiri, S.A.; Samsampour, D.; Seyahooei, M. A. and Bagheri, A. (2021). The effect of bacterial endophyte (*Exigubacterium aurantiacum*) isolated from salsola imbricata on growth characteristics of tomato seedlings under salinity stress. J. of Horti. Sci. 35(1): 153-167.
- De Pascale, S.; Maggio, A.; Fogliano, V.; Abrosino, P. and Ritieni, A. (2001). Irrigation with saline water improves carotenoids content and antioxidant activity of tomato. J. Hort. Sci Biotech 76: 447–453.
- De Pascale, S.; Ruggiero, C. and Barbieri, G. (2003). Physiological responses of pepper to salinity and drought. Soc. Hort. Sci. 128: 48–54.
- Del Amor, F.M.; Martinez, V. and Cerda, A. (2001). Salt tolerance of tomato plants as affected by stage of plant development. Hort. Sci. 36(7): 1260–1263.
- Desai, S.; Bagyaraj, D.J. and Revanna, A. (2020). Inoculation with microbial consortium promotes growth of tomato and capsicum seedlings raised in pro trays. Proceedings of the National Academy of Sciences, India -Section B: Biol. Sciences 90: 21-28.
- Dodd, I.C., Belimov, A.A., Sobeih, W.Y., Safronova, V.I., Grier-son, D., and Davies, W.J. (2004). Will modifying plant ethylenestatus improve plant productivity in

water-limited environments? In Proceedings of the 4<sup>th</sup> International Crop Science Congress, Brisbane, Australia, 26 September - 1 October 2004. Edited by T. Fischer, N. Turner, J. Angus, L. McIntyre, M. Robertson, A.Borrell, and A. Lloyd. The Regional Institute Ltd., Gosford, NSW, Australia. 1-5.

- Du Jardin, P. (2015). Plant biostimulants: defnition, concept, main categories and regulation. Sci. Hort. 196: 3–14.
- Dursun, A.; Guvenc, I. and Turan, M. (2002). Effects of different levels of humic acid on seedling growth and macro and micronutrient contents of tomato and eggplant. Acta Agrobotanica. 56: 81-88.
- Ebrahim, M.K.H. and Saleem, A.R. (2017).
  Alleviating salt stress in tomato inoculated with mycorrhizae: Photosynthetic performance and enzymatic antioxidants. J. of Taibah Univ. for Sci. 850-860.
- El-Galad, M.A.; Sayed, D. and El-Shal, R.M. (2013). Effect of humic acid and compost applied alone or in combination with sulphur on soil fertility and faba bean productivity under saline soil conditions. J. Soil Sci. and Agric. Eng., Mansoura Univ. 4 (10): 1139 1157.
- El-Mogy, M. M.; Garchery, C. and Stevens, R. (2018). Irrigation with salt water affects growth, yield, fruit quality, storability and marker-gene expression in cherry tomato. Acta Agriculturae Scandinavica, Section B Soil & Plant Sci. 68: 727-737.
- Eraslan, F.; Inal, A.; Gunes, A.; Alpaslan, M. and AtIkmen, N. C. (2015). Comparative physiological and growth responses of tomato and pepper plants to fertilizer induced salinity and salt stress. Fresenius Environmenttal. Bulletin. 24(5a): 1774-1778.
- Evelin, H.; Giri, B. and Kapoor, R. (2012). Contribution of *Glomus intraradices* inoculation to nutrient acquisition and mitigation of ionic imbalance in NaCl-stressed *Trigonella foenum-graecum*. Mycorrhiza. 22: 203–217.
- Feigin, A.; Rylski, I.; Meiri, A. and Shalhevet, J. (1987). Response of melon and tomato plants

to chloride-nitrate ratio in saline nutrient solution. J. Plant Nutr. 10: 1787-1794.

- Feleafel, M. N. and Mirdad, Z. M. (2014a). Alleviating the deleterious effects of water salinity on greenhouse grown tomato. International J. of Agri. and Biology. 16(1): 49-56.
- Feleafel, M. N. and Mirdad, Z. M. (2014b). Ameliorating tomato productivity and wateruse efficiency under water salinity. JAPS, J.of Animal and Plant Sci. 24: 302-309.
- Friedel, J.K. and Scheller, E. (2002). Composition of hydrolysable amino acids in soil organic matter and soil microbial biomass. Soil Biol. Biochem. 34: 315–325.
- Gama, P.B.; Inanaga, S.; Tanaka, K. and Nakazawa, R. (2007). Physiological response of common bean (*Phaseolus vulgaris*) seedlings to salinity stress. Afr. J. Biotech. 6: 79-88.
- Garg, N. and Chandel, S. (2010). Arbuscular mycorrhizal network: process and functions A review. Agron. Sustain. Develop. 39: 581– 599.
- Glick, B.R. (2012). Plant Growth-Promoting Bacteria: Mechanisms and Applications. Scientifica.18:1-15.
- Glick, B.R. (2014). Bacteria with ACC deaminase can promote plant growth and help to feed the World. Microbiol. Res. 169: 30–39.
- Grattan, S.R. and Grieve, C.M. (1999). Salinity mineral nutrient relations in horticultural crops. Sci. Hort., 78: 127 – 157.
- Greenway, H. and Munns, R. (1980). Mechanisms of salt tolerance in nonhalophytes. Annu. Rev. Plant Physiol., 31: 149 - 190.
- Hadad, M.A.; Al-Hashmi, H. S. and Mirghani, S. M. (2012). Tomato (*Lycopersicon esculentum Mill.*) growth in response to salinity and inoculation with native and introduced strains of mycorrhizal fungi. Inter.Res. J. of Agric. Sci. and Soil Sci. 2: 228-233.
- Hajiboland, R.; Aliasgharzadeh, N.; Laiegh, S. F. and Poschenrieder, C. (2010). Colonization with arbuscular mycorrhizal fungi improves salinity tolerance of tomato (*Solanum*)

*lycopersicum L.*) plants. Plant Soil 331: 313–327.

- Hammad, S.A.; Shaban, K.H. A. and Tantawy, M.F. (2010). Studies on salinity tolerance of two peanut cultivars in relation to growth, leaf water content. Some chemical aspect and yield. J. Appl. Sci. Res. 6: 1517 – 1526.
- Hashem, A.; Tabassum, B. and Abd Allah, E, F. (2019). Bacillus subtilis: A plant-growth promoting rhizobacterium that also impacts biotic stress. Saudi J. of Biol. Sci. 1291-1297.
- Heikham, E.; Kapoor, R. and Giri, B. (2009). Arbuscular Mycorrhiza fungi in alleviation of salt stress: a review. Ann. Bot., 104: 1263.
- Helaly, A.A.; Goda, Y. and Abd EL-Rehim, A.S. (2017). Effect of irrigation with different levels of saline water type on husk tomato productivity. Adv. Plants Agric. Res. 6:1-8.
- Ho, L.C.; Zamski, E. and Schaffer, A.A. (1996). Photoassimilate distribution in plants and crops. Sink Relationships, Marcel Dekker, Inc, 709-728.
- Huang, J.C.; Lai, W.A.; Singh, S.; Hameed, A. and Young, C.C. (2013). Response of mycorrhizal hybrid tomato cultivars under saline stress. J. of Soil Sci. and Plant Nutr. 13: 469-484.
- Isfahani, F. M.; Tahmourespour, A.; Hoodaji, M.; Ataabadi, M. and Mohammadi, A. (2019).
  Influence of exopolysaccharide-producing bacteria and SiO<sub>2</sub> nanoparticles on proline content and antioxidant enzyme activities of tomato seedlings (*Solanum lycopersicum L.*) under salinity stress. Polish J. of Environ. Studies. 28: 153-163.
- Jahromi, F.; Aroca, R.; Porcel, R. and Ruiz-Lozano, J.M. (2008). Influence of salinity on the in vitro development of *Glomus intraradices* and on the in vivo physiological and molecular responses of mycorrhizal lettuce plants. Micro. Eco. 55: 45–53.
- James, R.A.; Blake, C.; Byrt, C.S. and Munns, R. (2011). Major genes for Na<sup>+</sup> exclusion, Nax1 and Nax2 (wheat HKT1; 4 and HKT1; 5), decrease Na+ accumulation in bread wheat leaves under saline and waterlogged conditions. J. of Exp. Botany. 62: 2939-2947.

Alleviation salinity stress on tomato plants by some organic and bio-fertilizers application

- Jianmin, Y.; Smith, M. D.; Glick, B. R. and Yan, L. (2014). Effects of ACC deaminase containing rhizobacteria on plant growth and expression of Toc GTPases in tomato (*Solanum lycopersicum*) under salt stress. Botany. 92: 775-781.
- Johanson, C.M. and Ulrichs, A. (1959). Analytical method for use in plant analysis, U.S. Dept. Agric. Inform. Bull.766.
- Johnson, R.W.; Dixon, M.A. and Lee, D.R. (1992). Water relations of the tomato during fruit growth. Plant, Cell Environ. 15: 947–953.
- Karlidag, H.; Yildirim, E.; Turan, M.; Pehluvan, M. and Donmez, F. (2013). Plant Growthpromoting rhizobacteria mitigate deleterious effects of salt stress on strawberry plants (*Fragaria ×ananassa*). Hortscience. 48: 563– 567.
- Kaya, C.; Ashraf, M.; Sonmez, O.; Aydemir, S.; Tuna, A.L. and Cullu, M.A. (2009). The influence of arbuscular mycorrhizal colonisation on key growth parameters and fruit yield of pepper plants grown at high salinity. Sci. Hort. 121:1–6.
- Knicker, H.; R. Frund and Ludemann, H.D. (1993). The chemical nature of nitrogen in native soil organic matter. Nat. Wissenschaften, 80: 219–221.
- Kohler, J.; Herna´ ndez, J. A.; Caravaca, F. and Rolda´ n, A. (2009). Induction of antioxidant enzymes is involved in the greater effectiveness of a PGPR versus AM fungi with respect to increasing the tolerance of lettuce to severe salt stress. Environ. Exp. Bot. 65: 245– 252.
- Koide, R. (1993). Physiology of the mycorrhizal plant. Adv. Plant Pathol. 9:33-54.
- Kumar, U.; Gulati, I.J.; Rathiya, G.R. and Singh, M.P. (2017). Effect of saline water irrigation, humic acid and salicylic acid on soil properties, yield attributes and yield of tomato (*Lycopersicon esculentum* Mill.). Environment & Ecology 35 (1B): 449-453.
- Leogrande, R.; Lopedota, O.; Vitti, C.; Ventrella, D. and Montemurro, F. (2016). Saline water and municipal solid waste compost

application on tomato crop: effects on plant and soil. J.of Plant Nutr. 39(4): 491-501.

- Liang, Y.C.; Si, J.; Nikolic, M.; Peng, Y.; Chen, W. and Jiang, Y. (2005). Organic manure stimulates biological activity and barley growth in soil subject to secondary salinization. Soil Biol. Biochem. 37: 1185– 1195.
- Magan, J.J.; Gallardo, M.; Thompson, R.B. and Lorenzo, P. (2008). Effects of salinity on fruit yield and quality of tomato grown in soil-less culture in greenhouses in Mediterranean climatic conditions. Agric Water Manag. 95: 1041–1055.
- Maggio, A.; De Pascale, S.; Angelino, G.; Ruggiero, C. and Barbieri, G. (2004).
  Physiological response of tomato to saline irrigation in long-term salinized soils. Europ.
  J. Agron. 21: 149-159.
- Malash, N. M.; Ali, F. A.; Fatahalla, M. A.; Khatab, Entsar. A.; Hatem, M. K. and Tawfic, S. (2008). Response of tomato to irrigation with saline water applied by different irrigation methods and water management strategies. International J. of Plant Produc. 2: 101-116.
- Malash, N.M.; Ali, F.A.; Fatahalla, M.A. and Tawfic, S. (2012). Response of tomato to irrigation with saline water applied by different irrigation methods and water management strategies. Int J Plant Prod. 2: 101–116.
- Malash, N.M.; Ghaibeh, A.; Abdelkrim, G.; Yeo, A.R.; Flowers, T.J.; Ragab, R. and Cuartero, J. (2002). Effect of irrigation water salinity on yield and fruit quality of tomato. Acta Hort. 573: 423 -427.
- Manaa, A.; Ahmed, H. B.; Smiti, S. and Faurobert, M. (2011). Salt-stress induced physiological and proteomic changes in tomato (*Solanum lycopersicum*) seedlings. (Plant Stress - Special issue 1.) OMICS A J. of Integrative Biol. 15: 801-809.
- Mantelin, S. and Touraine, B. (2004). Plant growth-promoting bacteria and nitrate availability impacts on root development and nitrate uptake. J. Exp. Bot. 55: 27–34.

- Marulanda, A.; Azco'n, R. and Ruiz-Lozano, J.M. (2003). Contribution of six arbuscular mycorrhizal fungal isolates to water uptake by *Lactuca sativa* plants under drought stress. Physiol Planta. 119: 526-533.
- Marulanda, A.; Barea, J. M. and Azco´ n, R. (2006). An indigenous drought-tolerant strain of *Glomus intraradices* associated with a native bacterium improves water transport and root development in *Retama sphaerocarpa*. Microb. Ecol. 52: 670–678.
- Mayak, S.; Tirosh, T. and Glick, B. R. (2004a). Plant growth promoting bacteria that confer resistance to water stress in tomato and peppers. J. Plant Sci. 166: 525-530.
- Mayak, S.; Tirosh, T. and Glick, B.R. (2004b). Plant growth-promoting bacteria that confer resistance in tomato to salt stress. Plant Physiol. Biochem. 42: 565-572.
- Menary, R.C. and Staden, J.V. (1976). Effects of phosphorus nutrition and cytokinins on flowering in the tomato, *Lycopersicon esculentum Mill*. Aust. J. Plant Physiol. 3: 201–205.
- Miceli, A.; Moncada, A. and Vetrano, F. (2021). Use of microbial biostimulants to increase the salinity tolerance of vegetable transplants. Agronomy. 2: 25.
- Mizrahi, Y.; Taleisnik, E.; Kagan-Zur, V.; Zohas, Y.; Offenbach, R.; Matan, E. and Golan, R. (1988). A saline irrigation regime for improving tomato fruit quality without reducing yield. J. Am. Soc. Hort. Sci. 113: 202–205.
- Moghaddam, J. R.; Hosseini, Y.; Nikpour, M. R. and Abdoli, A. (2018). Evaluation the effects of the irrigation water salinity and water stress on yield components of cherry tomato. J. Water and Soil. 32(3): 489-500.
- Mohammad, M.; Shibli, R.; AJlouni, M. and Nimri, L. (1998). Tomato root and shoot responses to salt stress under different levels of phosphorus nutrition. J. of Plant Nutr. 21(8): 1667-1680.
- Mostafizur Rahman, M.D.; Hossain, M.; Kaniz Fatima Binte Hossain.; Sikder, M.D.T.; Shammi, M.; Rasheduzzaman, M.D.; Hossain,

M.A.; Alam, A.M. and Uddin, M.K. (2018). Effects of NaCl-salinity on tomato (*Lycopersicon esculentum Mill.*) plants in a pot experiment. Open Agric. 3: 578–585.

- Munns, R. and Tester, M. (2008). Mechanisms of salinity tolerance. Annual Rev. of Plant Biol. 59: 651–681.
- Niu, X.; Bressan, R.A.; Hasegawa, P.M. and Pardo, J.B. (1995). Ion homeostasis in NaCl stress environments. Plant Physiol. 109: 735– 742.
- Ors, S.; Ekinci, M.; Yildirim, E.; Sahin, U.; Turan, M. and Dursun, A. (2021). Interactive effects of salinity and drought stress on photosynthetic characteristics and physiology of tomato (*Lycopersicon esculentum L.*) seedlings. South Afric. J. of Botany. 137: 335-339.
- Oztekin, G.B. and Tuzel, Y. (2011). Comparative salinity responses among tomato genotypes and rootstocks. Pak. J. Bot. 43: 2665-2672.
- Oztekin, G.B.; Tuzel, Y. and Tuzel, I.H. (2013). Does mycorrhiza improve salinity tolerance in grafted plants? Scientia Horticulturae.149: 55-60.
- Padem, H.; Ocal, A. and Alan, R. (1997). Effect of humic acid added foliar fertilizer on seedling quality and nutrient content of eggplant and pepper. ISHS Symposium on Greenhouse Management for Better Yields and Quality in Mild Winter Climates. Acta Horti. 491: 241-246.
- Page, A. L.; Miller, R.H. and Keeney, D. R. (1982). (Methods of soil analysis). II. Chemical and Microbiogical properties 2<sup>nd</sup> Ed. Madison, Wisconsim. U.S.A.
- Pandey, S. and Gupta, S. (2020). Evaluation of *Pseudomonas sp.* for its multifarious plant growth promoting potential and its ability to alleviate biotic and abiotic stress in tomato (*Solanum lycopersicum*) plants. Scientific Reports. 10: 1-15.
- Pengfei, Z., Yan, D.Y., Masateru, S.and Natsumi, M. (2019). Interactive influences of salinity stress and leaf thinning on the growth, yield, and water use efficiency, and fruit quality of

cherry tomatoes. Communications in Soil Sci. and Plant Anal. 50:1003-1012.

- Pergel, F. (1945). Quantitative organic micro. Analysis, 4<sup>th</sup> Ed. J. &A. Chuch. II, I td. London.
- Petreikov, M.; Yeselson, Y.; Shen, S.; Levin, I.; Schaffer, A.; Efrati, A. and Moshe, B. (2009). Carbohydrate balance and accumulation during development of near-isogenic tomato lines differing in the AGPase-L1 allele. J. Am. Soc. Hort. Sci. 134: 134-140.
- Pietrantonio, L.; Golubkina, N. A.; Cozzolino, E.; Sellitto, M.; Cuciniello, A. and Caruso, G. (2020). Yield and quality performances of tomato "plum" inoculated with arbuscular mycorrhizal fungi in saline soils. Acta Horticulturae. (1271): 351-357.
- Psarras, G.; Bertaki, M. and Chartzoulakis, K. (2008). Response of greenhouse tomato to salt stress and K<sup>+</sup> supplement. Plant Biosyst. 142: 149–153.
- Qadir, M. and Oster, J.D. (2004). Crop and irrigation management strategies for salinesodic soils and waters aimed at environmentally sustainable agriculture. Sci. Total Environment. 323: 1–19.
- Radwan, A.A.; Hassan, A.A. and Malash, N.M. (1979). Physiological studies on tomato fruits firmness, total soluble solids and vitamin C contents. Res. Bull. No. 1063, Faculty of Agric. Ain Shams University. Hort. Abstr., 50: 9141, 1980).
- Rady, M. M. (2012). A novel organo-mineral fertilizer can mitigate salinity stress effects for tomato production on reclaimed saline soil. South Afric. J. of Botany. 81: 8-14.
- Rady, M.M.; Semida, W.M.; Hemida, K.A. and. Abdelhamid, M.T. (2016). The effect of compost on growth and yield of *Phaseolus vulgaris* plants grown under saline soil. Int. J. Recycl Org Waste Agric. 5: 311–321.
- Rahnama, A.; James, R.A.; Poustini, K. and Munns, R. (2010). Stomatal conductance as a screen for osmotic stress tolerance in durum wheat growing in saline soil. Functional Plant Biol. 37:255-263.

- Rani, P.; Sharma, M. K.; Rani, S.; Kumar, N. and Sharma, S. K. (2017). Effect of different saline environments on yield and quality of tomato (*Lycopersicon esculentum L.*). Annals of Agri. Bio. Res. 22(2): 223-227.
- Richardson, A. E.; Barea, J. M.; McNeill, A. M. and Prigent-Combaret, C. (2009). Acquisition of phosphorus and nitrogen in the rhizosphere and plant growth promotion by microorganisms. Plant and Soil. 321: 305– 339.
- Rozema, J. and Flowers, T. (2008). Crops for a Salinized World. Sci. 322, 1478-1480.
- Ruiz-Lozano, J.M. and Azcon, R. (1995). Hyphal contribution to water uptake in mycorrhizal plants as affected by the fungal species and water status. Physiol. Plant. 95: 472-478.
- Saha, D.; Fakir, O.A.; Mondal, S. and Ghosh, R.C. (2017). Effects of organic and inorganic fertilizers on tomato production in saline soil of Bangladesh. J. Sylhet Agril. Univ., 4(2): 213-220.
- Saidimoradi, D.; Ghaderi, N. and Javadi, T. (2019). Salinity stress mitigation by humic acid application in strawberry (*Fragaria x ananassa Duch.*). Sci. Horti., 256: 1-15.
- Saito, T. and Ito, H. (1967). Studies on the growth and fruiting in tomato X. Effects of early environmental conditions and cultural treatments on the morphological and physiological development of flower and flower drop 2. Effect of watering, defoliation and application of gibberellin. J Jpn Soc Hortic Sci., 36: 281–289.
- Saranga, Y.; Zamir, D.; Marani, A. and Rudich, J. (1993). Breeding tomatoes for salt tolerance: variations in ion concentration associated with response to salinity. J. Amer. Hort. Sci. 118: 405–408.
- Schaffer, A.A. I.; Levin, I. I.; Ogus, I. M.; Petreikov, M.; Cincarevsky, F.; Yeselson, E.; Shen, S.; Gilboa, N. and Bar, M. (2000). ADPglucose pyrophosphorylase activity and starch accumulation in immature tomato fruit: The effect of a *Lycopersicon hirsutum*-derived introgression encoding for the large subunit. Plant Sci. 152: 135-144.

- Scholberg, J.M.S. and Locascio, S.J. (1999). Growth response of snap bean and tomato as affected by salinity and irrigation method. Hort Sci. 34 (2): 259-264.
- Semida, W. M.; El-Mageed, T. A. A. and Howladar, S. M. (2014). A novel organomineral fertilizer can alleviate negative effects of salinity stress for eggplant production on reclaimed saline calcareous soil. Acta Horti. (1034): 493-499.
- Shen, M.; Kang, Y.J.; Wang, H.L.; Zhang, X.S. and Zhao, Q.X. (2012). Effect of plant growthpromoting rhizobacteria (PGPR) on plant growth, yield, and quality of tomato (*Lycopersicon esculentum Mill.*) under simulated seawater irrigation. J. Gen. Appl. Microbiol. 58: 253–262.
- Snedecor, G. W. and W. G. Cochran (1980). Statistical Methods. 7<sup>th</sup> ED. Ames: Iowa State University Press.
- Soliman, M.S. and Doss, M. (1992). Salinity and mineral nutrition effects on growth and accumulation of organic and inorganic ions in two cultivated tomato varieties. Jour. of Plant Nut.
- Souza, A.C.; Zandonadi, D.B.; Santos, M.P.; Canellas, N.O.A.; Soares, C.D.P.; Irineu, L.E.S.D.S.; Rezende, C.E.D.; Spaccini, R.; Piccolo, A.; Olivares, F.L. and Canellas, L.P. (2021). Acclimation with humic acids enhances maize and tomato tolerance to salinity. Chem. Biol. Technol. Agric. 8 (40): 1-13.
- Souza, W.S. (1990). Produção e desenvolvimento do tomate industrial (Lycopersicun esculentum Mill) em diferentes níveis de salinidade. Campina Grande: UFPB, 65p. (Dissertação - Mestrado).
- Sun, X.C.; Hu, C.X. and Tan, Q.L. (2006). Effect of molybdenum on antioxidative defense system and membrane lipid peroxidation in winter wheat under low temperature stress. J. of Plant Physiology and molecular biology, 32: 175-182.
- Tan, K.H. (1998). Colloidal chemistry of organic soil constituents. In: Principles of Soil Chem.,

pp: 177–258. Tan, H. (ed.). Marcel Dekker, New York, USA.

- Tank, N. and Saraf, M. (2010). Salinity-resistant plant growth promoting rhizobacteria ameliorates sodium chloride stress on tomato plants. Plant Interactions. 5, No. 1: 51-58.
- Tartoura, K. A. H.; Youssef, S. A. and Tartoura, E. A. A. (2014). Compost alleviates the negative effects of salinity via up-regulation of antioxidants in *Solanum lycopersicum L*. plants. Plant Growth Regulation. 74: 299-310.
- Tejada, M.; Garcia, C.; Gonzalez, J.L. and Hernandez, M.T. (2006). Use of organic amendment as a strategy for saline soil remediation: influence on the physical, chemical and biological properties of soil. Soil Biol Biochem. 38: 1413–1421.
- Tester, M. and Davenport, R. (2003). Na<sup>+</sup> tolerance and Na<sup>+</sup> transport in higher plants. Annul. of Botany. 91: 303–327.
- Torre-Gonzalez, A. de La.; Montesinos-Pereira, D.; Blasco, B. and Ruiz, J. M. (2018). Influence of the proline metabolism and glycine betaine on tolerance to salt stress in tomato (*Solanum lycopersicum L.*) commercial genotypes. J. of Plant Physiol. 231: 329-336.
- Tu, C.; Ristaino, J.B. and Hu, S. (2006). Soil microbial biomass and activity in organic tomato farming systems: Effects of organic inputs and straw mulching, Soil Biol. Biochem. 38: 247–255.
- Turan, M.; Yildirim, E.; Ekinci, M. and Argin, S. (2021). Effect of biostimulants on yield and quality of cherry tomatoes grown in fertile and stressed soils. HortScience. 56(4): 414-423.
- Ullah, U.; Ashraf, M.; Shahzad, S.M.; Siddiqui, A.R.; Piracha, M.A. and Suleman, M. (2016). Growth behavior of tomato (Solanum lycopersicum L.) under drought stress in the presence of silicon and plant growth promoting rhizobacteria. Soil Environ. 35(1): 65-75.
- Vessey, J.K. (2003). Plant Growth Promoting Rhizobacteria as biofertilizers. Plant Soil .255: 571–586.

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- Viol, M. A., Carvalho, J. de A., Lima, E. M. de C., Rezende, F. C.and Gomes, L. A. A.(2017). Salinity effects of growth and production of tomato cultivated in greenhouse. Revista Brasileira de Agricultura Irrigada. 11: 2120-2131.
- Walker, D. J. and Bernal, M. P. (2008). The effects of olive mill waste compost and poultry manure on the availability and plant uptake of nutrients in a highly saline soil. Bioresource Technol. 99(2): 396-403.
- Wang, L.; Sun, X.; Li, S.; Zhang, T.; Zhang, W. and Zhai, P. (2014). Application of organic amendments to a coastal saline soil in north China: effects on soil physical and chemical properties and tree growth. PLoS ONE 9: 1-9.
- Wright, A.L.; Provin, TL; Hons, F.M.; Zuberer, D.A. and White, R.H. (2008). Compost impacts on sodicity and salinity in a sandy loam turf grass soil. Compost Science & Utilization. 16: 30–35.
- Yildirim, E.; Taylor, A.G. and Spittler, T.D. (2006). Ameliorative effects of biological treatments on growth of squash plants under salt stress. Sci. Hort., 111: 1–6.
- Yilmaz, Y.; Erdinc, C.; Akkopru, A. and Kipcak, S. (2020). Use of plant growth promoting rhizobacteria against salt stress for tomato

(*Solanum lycopersicum L.*) seedling growth. Acta Sci. Pol. Hortorum Cultus. 19: 15–29.

- Yurtseven, E.; Kesmez, G.D. and Unlukara, A. (2005). The effects of water salinity and potassium levels on yield, fruit quality and water consumption of a native central anatolian tomato species (*Lycopersicon esculantum*). Agric. Water Management. 78: 128-135.
- Zhang, P.; Senge, M. and Dai, Y. (2016). Effect of salinity stress on growth, yield, fruit quality and water use efficiency of tomato under hydroponics system. Agric. Sci. 4: 46-55.
- Zhai, Y.; Qian, Y. and MaoMao, H. (2015). The effects of saline water drip irrigation on tomato yield, quality, and blossom-end rot incidence - A 3a case study in the south of China. PLoS ONE.10: 1-17.
- Zhai, Y.; Qian, Y. and Yu, W.Y. (2016). Soil salt distribution and tomato response to saline water irrigation under straw mulching. PLoS ONE. 11: 1-17.
- Zribi, W.; Faci, J.M. and Aragues, R. (2011). Mulching effects on moisture, temperature, structure and salinity of agricultural soils. Información Técnica Económica Agraria. 107: 148-162.

### تخفيف إجهاد الملوحة على نباتات الطماطم من خلال بعض تطبيقات الأسمدة العضوية والحيوية

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#### الملخص العربى

أجريت هذة التجربة في موسمين زراعين متتالين ٢٠٢٠ و ٢٠٢١ في أصص تحت الصوبة السيرام بمزرعة التجارب بكلية الزراعة - جامعة المنوفية - بشبين الكوم. الهدف من هذه الدراسة هو دراسة تأثير التسميد العضوي (الكمبوست والهيوميك اسيد) والتسميد الحيوي (الميكورايزا والبكتريا المنشطة للنمو) لتخفيف الاثار السلبية للملوحة على هجين الطماطم ١٨٦. تم تجهيز محاليل ملحية بإستخدام كلوريد الصوديوم بتركيزين (٣ و٦ dS/m<sup>-1</sup>) بالاضافة الى الري بماء الصنبور ككنترول. أظهرت النتائج أن الملوحة أدت الى نقص في كل من: صفات النمو الخضري مثل إرتفاع النبات والوزن الجاف للنبات كذلك حدث نقص في عدد الايام اللازمة لبداية تز هير ٥٠٪ من النباتات ( F50) وكذا النسبة المئوية لعقد الثمار، المحتوى المائي النسبي في الاوراق، محتوى الاوراق الحديثة والمسنة من النيتر وجين والفوسفور والبوتاسيوم والكالسيوم بالاضافة الى انخفاض محصول الثمار ومكوناتة. وعلى الجانب الاخر أدت الملوحة الى زيادة في كل من : كفاءة استخدام الماء، محتوى الأور اق من البرولين ، التسريب من الجدر الخلوية في الاور اق بالاضافة الى تحسين جودة الثمار حيث ادت الملوحة الى زيادة محتوي الثمار من المواد الصلبة الذائبة الكلية (TSS) وفيتامين ج ،كما ادت الملوحة الى زيادة محتوي الاور اق الحديثة والمسنة من عنصري الصوديوم والكلور ولكن كان التركيز أعلى في الاور اق القديمة عن الحديثة. كما أدت معاملات التسميد الحيوي والعضوي (سواء استخدمت فردية أومجمعة ثنائية؛ وهي الكمبوست + الهيوميك اسيد معا وأيضا الميكورايزا + البكتريا المنشطة للنمومعا) الي تخفيف التأثير الضار للملوحة حيث أدت الى زيادة قيم كل من قياسات النمو الخضري ونسبة العقد كما أدت الى تحسين الحالة المائية للنبات وكفاءة استخدام الماء ومحتوي الاوراق الحديثة والمسنة من النيتر وجين والفوسفور والبوتاسيوم والكالسيوم والمحصول ومكوناتة وبالنسبة لصفات جودة الثمار فقد أدت معاملات تخفيف حدة الملوحة (الحيوية والعضوية) الى تقليل محتوى الثمار من المواد الصلبة الذائبة الكلية (TSS) في حين أدت الي زيادة محتوي الثمار من فيتامين C. معاملات تخفيف الاثار الضارة للملوحة (التسميد الحيوي والعضوي) أدت الى تقليل محتوي الاوراق الحديثة والمسنة من الصوديوم والكلور ،محتوي الاوراق من البرولين والتسريب في الاوراق مما أدي الى تحسين النمو والانتاجية لنباتات الطماطم. المعاملات المجمعة (الميكور ايزا مع البكتريا المنشطة للنمو) و(الكمبوست مع الهيوميك أسيد) كان لها تأثير تآزري حيث أدت الى تخفيف تأثير الملوحة بدرجة افضل من استخدام كل مكون على حدة. وعلى هذا فقد أدت هذة المعاملات المجمعة (الثنائية) الى أفضل النتائج تلاها المعاملة بكل من الكمبوست والميكور ايزا (والتي استخدمت بمفردها) من حيث تخفيف التأثير الضار للملوحة على نباتات الطماطم.

**الكلمات المفتاحية:** الطماطم ، معاملات تخفيف الملوحة <sup>،</sup> نمو النبات ، المحتويات الكيميائية ومحصول الثمار ، التسميد العضوي والحيوي.

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